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(54) Title: NEUROPEPTIDE Y RECEPTOR YS AND NUCLEIC ACID SEQUENCE

(57) Abstract

The present invention provides novel NPY/PYY receptor proteins and the nucleic acid sequence encoding them. The invention is directed to the isolation, characterization, and pharmacological use of these receptors and nucleic acids. In particular, this invention provides human and rat NPY/PYY receptors (which we call the NPY Y5 receptor) and nucleic acids. Also provided are recombinant expression constructs useful for transfecting cells and expressing the protein in vitro and in vivo. The invention further provides methods for detecting expression levels of the protein as well as methods for screening for receptor antagonists and agonists to be used for the treatment of obesity or anorexia, respectively.

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Jonathan A. Bard, et al. U.S. Serial No.: 08/495,695 Filed: January 13, 1997 Exhibit 18

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NEUROPEPTIDE Y RECEPTOR Y5 AND NUCLEIC ACID SEQUENCE BACKGROUND OF THE INVENTION

Field of the Invention

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This invention relates to a novel neurotransmitter Neuropeptide Y receptor, its nucleic acid sequence, and compounds, compositions, and methods for their use.

Summary of the Related Art

Neuropeptide Y (NPY) is a 36-amino acid peptide neurotransmitter that is located throughout the central and peripheral nervous systems. Tatemoto, *Proc. Natl. Acad. Sci. USA* 79, 5485 (1982); Hazlewood, *Proc. Soc. Exp. Biol. Med.* 202, 44 (1993). It affects a broad range of phenomena, including blood pressure regulation, memory, anxiolysis/sedation, food and water appetite, vascular and other smooth muscle activity, intestinal electrolyte secretion, and urinary sodium excretion. *E.g.*, Colmers and Wahlestedt, *The Biology of Neuropeptide Y and Related Peptides* (Humana Press, Totowa, NJ, 1993); Kalra et al., *Phys. & Behavior* 50, 5 (1991).

Peptide YY (PYY) is also a 36 amino acid peptide and has significant sequence homology (70%) to NPY. Taternoto et al., Nature 296, 659 (1982). Its anatomical distribution is similar to that of NPY, although it is located mainly in the endocrine cells of the lower gastrointestinal tract. Bottcher et al., Regul. Pept. 8, 261 (1984). Like NPY, PYY stimulates feeding in rats. Morley et al., Brain Res. 341, 200 (1985). Along with the pancreatic polypeptide (PP), NPY and PYY have a common tertiary structure, characterized by the so-called PP-fold. Glover, Eur. J. Biochem. 142, 379 (1985). Both NPY and PYY show about a 50% sequence homology with PP.

Because of their structural similarities, NPY and PYY have a number of common receptors. At least four receptor subtypes, Y1, Y2, Y3, and Y4/PP, have been identified. The affinity for NPY, PYY, and various fragments thereof varies among the subtypes. See, e.g., Bard et al. (WO 95/17906) and references cited therein. For example, Y1 and Y2 subtypes have high affinity for NPY and PYY. Whereas Y1 has high affinity for (Leu³¹Pro³⁴)NPY ((LP)NPY)and low affinity for (13-36)NPY, Y2 behaves oppositely. Y3 has high affinity for NPY but low affinity for PYY. Y4/PP has a high affinity for PP but relatively low affinity for NPY.

Wahlestedt (WO 93/24515) and Larhammar et al. (J. Biol. Chem. 267, 10935 (1992)) describe the cloning and identification of the human Y1-type NPY/PYY receptor isolated from

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human fetal brain tissue. Selbie et al. (WO 93/09227) disclosed the full length cDNA sequence of the Y1 receptor from human hippocampus. Eva et al. (FEBS Lett. 271, 81 (1990)) cloned the NPY Y1 receptor from rat forebrain. Eva et al. (FEBS Lett. 314, 285 (1992)) cloned the NPY Y1 receptor from murine genomic DNA.

The Y2-type receptor has also been cloned. Gerald et al. (WO 95/21245) disclosed the cDNA sequence of human hippocampal Y2 and two rat Y2 clones. Rose et al. (J. Biol. Chem. 270, 22661 (1995)) disclosed the cDNA sequence of the Y2 receptor from a human neuroblastoma cell line.

Bard et al. (supra) and Lundell et al. (J. Biol. Chem. 270, 29123 (1995)) described cloning the cDNA sequence of the Y4/PP receptor from both rat spleen and human placenta.

To date, the Y3 receptor has not been cloned.

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Because of the important role of NPY and PYY in a number of physiological processes, such as feeding, there is a strong need to further develop materials and methods for investigating the mechanistic behavior of these compounds and for treating diseased and other abnormal states associated with the physiological processes in which NPY and PYY act. Specifically, the NPY analogs/fragments that induce feeding, such as (LP)(3-36)NPY, do not bind to the previously identified NPY/PYY receptors with affinities consistent with the feeding response. Accordingly, there is a need and desire to identify the NPY/PYY receptor that is responsible for the feeding response. Antagonists to such a receptor could be used to treat obesity and diabetes by reducing appetite and food consumption.

SUMMARY OF THE INVENTION

The present invention provides, inter alia, novel NPY/PYY receptor proteins. Also provided are the nucleic acid sequences encoding these novel receptor proteins, as well as compounds and methods for using these proteins and their nucleic acid sequences.

The present invention provides novel proteins, nucleic acids, and methods useful for developing and identifying compounds for the treatment of such diseases and disorders as obesity. Identified and disclosed herein is the protein sequence for a novel receptor for the neurotransmitters Neuropeptide Y (NPY) and Peptide YY (PYY) and the nucleic acid sequence encoding this receptor, which we call the NPY Y5 (or simply "Y5") receptor. The importance of this discovery is manifested in the effects of NPY, which include blood pressure regulation, memory enhancement, anxiolysis/sedation, and increased food intake. Thus, this receptor

protein is useful for screening for NPY/PYY agonist and antagonist activity for controlling these conditions.

In one aspect of the present invention, we provide isolated nucleic acid sequences for a novel NPY and PYY receptor, the Y5 receptor. In particular, we provide the cDNA sequences encoding for the rat and human receptors and isoforms thereof. These nucleic acid sequences have a variety of uses. For example, they are useful for making vectors and for transforming cells, both of which are ultimately useful for production of the Y5 receptor protein. They are also useful as scientific research tools for developing nucleic acid probes for determining receptor expression levels, e.g., to identify diseased or otherwise abnormal states. They are useful for developing analytical tools such as antisense oligonucleotides for selectively inhibiting expression of the receptor gene to determine physiological responses.

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In another aspect of the present invention, we provide a homogenous composition comprising the receptor Y5 protein. The protein is useful for screening drugs for agonist and antagonist activity, and, therefore, for screening for drugs useful in regulating physiological responses associated with the Y5 receptor. Specifically, antagonists to the Y5 receptor could be used to treat obesity and diabetes by reducing appetite and food consumption, whereas agonists could be used for the treatment of anorexic conditions. The proteins are also useful for developing antibodies for detection of the protein.

Flowing from the foregoing are a number of other aspects of the invention, including (a) vectors, such as plasmids, comprising the receptor Y5 nucleic acid sequence that may further comprise additional regulatory elements, e.g., promoters, (b) transformed cells that express the Y5 receptor, (c) nucleic acid probes, (d) antisense oligonucleotides, (e) agonists, (f) antagonists, and (g) transgenic mammals. Further aspects of the invention comprise methods for making and using the foregoing compounds and compositions.

The foregoing merely summarizes certain aspects of the present invention and is not intended, nor should it be construed, to limit the invention in any manner. All patents and other publications recited herein are hereby incorporated by reference in their entirety.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 displays the competition curves of various peptides for [125I]PYY to Y5 receptor membranes transiently expressed in COS-7 cells.

Figure 2 displays saturation curves for specific binding of [125I]PYY to Y5 receptor membranes transiently expressed in COS-7 cells.

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DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention comprises, in part, a novel NPY/PYY receptor protein, the Y5 receptor. Particularly preferred embodiments of the Y5 receptor are those having an amino acid sequence substantially the same as SEQ ID NOs 2, 4, or 6. As used herein, reference to the Y5 receptor is meant as a reference to any protein having an amino acid sequence substantially the same as SEQ ID NOs 2, 4, or 6. The present invention also comprises the nucleic acid sequence encoding the Y5 protein, which nucleic acid sequences is substantially the same as SEQ ID NOs 1, 3, or 5. Receptors SEQ ID NOs 2 and SEQ ID NO 4 are rat Y5 receptors and appear to be allelic variations, with SEQ ID NO 4 the most commonly occurring and, therefore, the preferred embodiment of the rat Y5 receptor of this invention. SEQ ID NO 6 is the human Y5 receptor and its preferred embodiment.

As used herein, a protein "having an amino acid sequence substantially the same as SEQ ID NO x" (where "x" is the number of one of the protein sequences recited in the Sequence Listing) means a protein whose amino acid sequence is the same as SEQ ID NO x or differs only in a way such that IC₅₀[(3-36)NPY], IC₅₀[(Leu³¹Pro³⁴)NPY], and IC₅₀[(Leu³¹Pro³⁴)(3-36)NPY] as determined according to the method detailed in Example 4, *infra*, are less than or equal to 30 nM. The NPY fragments (3-36)NPY, (Leu³¹Pro³⁴)NPY and (Leu³¹Pro³⁴)(3-36)NPY induce a feeding response. Those skilled in the art will appreciate that conservative substitutions of amino acids can be made without significantly diminishing the protein's affinity for NPY, PYY, and fragments and analogs thereof. Other substitutions may be made that increase the protein's affinity for these compounds. Making and identifying such proteins is a routine matter given the teachings herein, and can be accomplished, for example, by altering the nucleic acid sequence encoding the protein (as disclosed herein), inserting it into a vector, transforming a cell, expressing the nucleic acid sequence, and measuring the binding affinity of the resulting protein, all as taught herein.

As used herein the term "a molecule having a nucleotide sequence substantially the same as SEQ ID NO y" (wherein "y" is the number of one of the protein-encoding nucleotide sequences listed in the Sequence Listing) means a nucleic acid encoding a protein "having an amino acid sequence substantially the same as SEQ ID NO y+1" (wherein "y+1" is the number of the amino acid sequence for which nucleotide sequence "y" codes) as defined above. This definition is intended to encompass natural allelic variations in the Y5 sequence. Cloned nucleic acid provided by the present invention may encode Y5 protein of any species of origin, including (but not limited to), for example, mouse, rat, rabbit, cat, dog, primate, and human. Preferably the nucleic acid provided by the invention encodes Y5 receptors of mammalian, and most preferably, rat or human origin.

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The invention also includes nucleotide sequences encoding chimeric proteins comprised of parts of the Y5 receptor and parts of other related seven-transmembrane receptors.

The 6B clone (SEQ ID NO 1) (see Example 2, infra) has a 2.4 kb cDNA insert with a open reading frame from nucleotide 248 to 1582 that encodes a 445 amino acid protein (SEQ ID NO 2). Hydrophobicity plot analysis using PEPPLOT of GCG shows that the Y5 receptor has seven transmembrane-like domains, indicating it might be a G-protein-coupled receptor. Unlike other known subtypes of NPY receptor family, the third intracellular loop of the Y5 receptor is unusually long. Another novel feature of the Y5 peptide sequence is that it has a much shorter C-terminal tail sequence than other known members of the NPY receptor family. It is also important to note that the Y5 sequence shows only 30-33% amino acid sequence identity to other NPY receptors.

Nucleic acid hybridization probes provided by the invention are DNAs consisting essentially of the nucleotide sequences complementary to any sequence depicted in SEQ ID NOs 1, 3, and 5 that is effective in nucleic acid hybridization. Nucleic acid probes are useful for detecting Y5 gene expression in cells and tissues using techniques well-known in the art, including, but not limited to, Northern blot hybridization, in situ hybridization, and Southern hybridization to reverse transcriptase - polymerase chain reaction product DNAs. The probes provided by the present invention, including oligonucleotide probes derived therefrom, are also useful for Southern hybridization of mammalian, preferably human, genomic DNA for screening for restriction fragment length polymorphism (RFLP) associated with certain genetic disorders. As used herein, the term complementary means a nucleic acid having a sequence that is sufficiently complementary in the Watson-Crick sense to a target nucleic acid to bind to the

target under physiological conditions or experimental conditions those skilled in the art routinely use when employing probes.

Receptor Y5 binds various fragments and analogs of NPY and PYY with affinities different from that of the known receptors. The rank order of binding affinity of receptor Y5 was found to be:

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NPY = (LP)NPY = PYY = (3-36)NPY = (LP)(3-36)NPY > (10-36)NPY > (18-36)NPYTable 1, *infra*, presents a more detailed affinity profile of the Y5 receptor for NPY, PYY, and various fragments thereof. As used herein, a protein having substantially the same affinity profile as the Y5 receptor means a protein in which the IC_{50} of each of the peptides listed in Table 1, *infra*, is no more than an order of magnitude greater than those listed in Table 1 for each of the respective peptides as measured according to the methods described in Example 4. Importantly, the NPY analogs/fragments that induce feeding, such as (LP)(3-36)NPY, do not bind to the previously identified NPY/PYY receptors with affinities consistent with the feeding response.

The production of proteins such as receptor Y5 from cloned genes by genetic engineering means is well known in this art. The discussion which follows is accordingly intended as an overview of this field, and is not intended to reflect the full state of the art.

DNA which encodes receptor Y5 may be obtained, in view of the instant disclosure, by chemical synthesis, by screening reverse transcripts of mRNA from appropriate cells or cell line cultures, by screening genomic libraries from appropriate cells, or by combinations of these procedures, as illustrated below. Screening of mRNA or genomic DNA may be carried out with oligonucleotide probes generated from the Y5 gene sequence information provided herein. Probes may be labeled with a detectable group such as a fluorescent group, a radioactive atom or a chemiluminescent group in accordance with known procedures and used in conventional hybridization assays, as described in greater detail in the Examples below. In the alternative, the Y5 gene sequence may be obtained by use of the polymerase chain reaction (PCR) procedure, with the PCR oligonucleotide primers being produced from the Y5 gene sequence provided herein. See U.S. Patent Nos. 4,683,195 to Mullis et al. and 4,683,202 to Mullis.

Receptor Y5 may be synthesized in host cells transformed with a recombinant expression construct comprising a nucleic acid encoding the receptor Y5. Such a recombinant expression construct can also be comprised of a vector that is a replicable DNA construct. Vectors are used herein either to amplify DNA encoding Y5 and/or to express DNA which

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encodes Y5. For the purposes of this invention, a recombinant expression construct is a replicable DNA construct in which a DNA sequence encoding Y5 is operably linked to suitable control sequences capable of effecting the expression of Y5 in a suitable host. The need for such control sequences will vary depending upon the host selected and the transformation method chosen. Generally, control sequences include a transcriptional promoter, an optional operator sequence to control transcription, a sequence encoding suitable mRNA ribosomal binding sites, and sequences which control the termination of transcription and translation. Amplification vectors do not require expression control domains. All that is needed is the ability to replicate in a host, usually conferred by an origin of replication, and a selection gene to facilitate recognition of transformants. See, Sambrook et al., Molecular Cloning: A Laboratory Manual (2nd Edition, Cold Spring Harbor Press, New York, 1989).

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Vectors useful for practicing the present invention include plasmids, viruses (including phage), retroviruses, and integratable DNA fragments (i.e., fragments integratable into the host genome by homologous recombination). The vector replicates and functions independently of the host genome, or may, in some instances, integrate into the genome itself. Suitable vectors will contain replicon and control sequences which are derived from species compatible with the intended expression host. The vectors may be self-replicating. Suitable vectors for the purposes of the present invention include pBluescript, pcDNA3, and, for insect cells, baculovirus. A preferred vector is the plasmid pcDNA3 (Invitrogen).

Construction of suitable vectors containing the desired coding and control sequences employs standard ligation and restriction techniques that are well understood in the art. Isolated plasmids, DNA sequences, or synthesized oligonucleotides are cleaved, tailored, and relegated in the form desired.

Site-specific DNA cleavage is performed by treating with the suitable restriction enzyme (or enzymes) under conditions that are generally understood in the art, and the particulars of which are specified by the manufacturer of these commercially available restriction enzymes. See, e.g., New England Biolabs, Product Catalog. In general, about 1 µg of plasmid or DNA sequence is cleaved by one unit of enzyme in about 20 µl of buffer solution. Often excess of restriction enzyme is used to ensure complete digestion of the DNA substrate. Incubation times of about one hour to two hours at about 37°C are workable, although variations are tolerable. After each incubation, protein is removed by extraction with phenol/chloroform, and may be followed by ether extraction. The nucleic acid may be recovered from aqueous fractions by

precipitation with ethanol. If desired, size separation of the cleaved fragments may be performed by polyacrylamide gel or agarose gel electrophoresis using standard techniques. A general description of size separations is found in *Methods in Enzymology* 65, 499-560 (1980).

Transformed host cells are cells which have been transformed or transfected with recombinant expression constructs made using recombinant DNA techniques and comprising mammalian Y5-encoding sequences. Preferred host cells for transient transfection are COS-7 cells. Transformed host cells may ordinarily express Y5, but host cells transformed for purposes of cloning or amplifying nucleic acid hybridization probe DNA need not express the receptor. When expressed, the mammalian Y5 protein will typically be located in the host cell membrane. See, Sambrook et al., ibid.

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Cultures of cells derived from multicellular organisms are desirable hosts for recombinant Y5 protein synthesis. In principal, any higher eukaryotic cell culture is workable, whether from vertebrate or invertebrate culture. However, mammalian cells are preferred, as illustrated in the Examples. Propagation of such cells in cell culture has become a routine procedure. See Tissuc Culture (Academic Press, Kruse & Patterson, Eds., 1973). Examples of useful host cell lines are bacteria cells, insect cells, yeast cells, human 293 cells, VERO and HeLa cells, LMTK cells, and WI138, BHK, COS-7, CV, and MDCK cell lines. Human 293 cells are preferred.

The invention provides homogeneous compositions of mammalian Y5 produced by transformed eukaryotic cells as provided herein. Such homogeneous compositions are intended to be comprised of mammalian Y5 protein that comprises at least 90% of the protein in such homogeneous composition. The invention also provides membrane preparation from cells expressing Y5 as the result of transformation with a recombinant expression construct, as described here.

Mammalian Y5 protein made from cloned genes in accordance with the present invention may be used for screening compounds for Y5 agonist or antagonist activity, or for determining the amount of a Y5 agonist or antagonist drug in a solution (e.g., blood plasma or serum). For example, host cells may be transformed with a recombinant expression construct of the present invention, Y5 protein expressed in those host cells, the cells lysed, and the membranes from those cells used to screen compounds for Y5 binding activity. Competitive binding assays in which such procedures may be carried out are well known in the art. By selection of host cells which do not ordinarily express Y5, pure or crude preparations of

membranes containing Y5 can be obtained. Further, Y5 agonists and antagonists can be identified by transforming host cells with a recombinant expression construct as provided by the present invention. Membranes obtained from such cells (and membranes of intact cells) can be used in binding studies wherein the drug dissociation activity is monitored.

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It is known that the neurotransmitter NPY is a regulator of appetite. As shown herein, the various NPY analogs/fragments that induce feeding, such as (LP)(3-36)NPY, bind with a high affinity to the Y5 receptor. Conversely, the NPY analogs/fragments that bind to the Y5 receptor with a lower affinity, such as (20-36)NPY, do not elicit feeding. It is therefore evident that by contacting the Y5 receptor with agonists and antagonists, feeding can be modulated. Accordingly, antagonists to the Y5 receptor, identified by the methods described herein, can be used to reduce appetite and hence treat obesity, diabetes and hyperlipidemia, and, conversely, agonists to the Y5 receptor can be used to treat conditions such as anorexia.

This invention provides a pharmaceutical composition comprising an effective amount of a agonist or antagonist drug identified by the method described herein and a pharmaceutically acceptable carrier. Such drugs and carrier can be administered by various routes, for example oral, subcutaneous, intramuscular, intravenous or intracerebral. The preferred route of administration would be oral at daily doses of about 0.01-100 mg/kg.

This invention provides a method of treating obesity, diabetes or hyperlipidemia, wherein the abnormality is improved by reducing the activity of Y5 receptor or blocking the binding of ligands to a Y5 receptor, which method comprises administering an effective amount of the antagonist-containing pharmaceutical composition described above to suppress the subject's appetite. Similarly, the invention also provides methods for treating diseases and conditions resulting from underfeeding and/or a loss of appetite, which method comprises administering an effective amount of an agonist-containing pharmaceutical composition described above to stimulate the subject's appetite.

The recombinant expression constructs of the present invention are useful in molecular biology to transform cells which do not ordinarily express Y5 to thereafter express this receptor. Such cells are useful as intermediates for making cell membrane preparations useful for receptor binding assays, which are in turn useful for drug screening. Drugs identified from such receptor assays can be used for the treatment of obesity, diabetes or anorexia.

The recombinant expression constructs of the present invention are also useful in gene therapy. Cloned genes of the present invention, or fragments thereof, may also be used in gene

therapy carried out by homologous recombination or site-directed mutagenesis. See generally Thomas & Capecchi, Cell 51, 503-512 (1987); Bertling, Bioscience Reports 7, 107-112 (1987); Smithies et al., Nature 317, 230-234 (1985).

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Oligonucleotides of the present invention are useful as diagnostic tools for probing Y5 gene expression in tissues. For example, tissues are probed in situ with oligonucleotide probes carrying detectable groups by conventional autoradiographic techniques, as explained in greater detail in the Examples below, to investigate native expression of this receptor or pathological conditions relating thereto. Further, chromosomes can be probed to investigate the presence or absence of the Y5 gene, and potential pathological conditions related thereto, as also illustrated by the Examples below. Probes according to the invention should generally be at least about 15 nucleotides in length to prevent binding to random sequences, but, under the appropriate circumstances may be smaller.

The invention also provides antibodies that are immunologically reactive to a mammalian Y5, preferably rat or human Y5. The antibodies provided by the invention are raised in animals by inoculation with cells that express a mammalian Y5 or epitopes thereof, using methods well known in the art. Animals that are used for such inoculations include individuals from species comprising cows, sheep, pigs, mice, rats, rabbits, hamsters, goats and primates. Preferred animals for inoculation are rodents (including mice, rats, hamsters) and rabbits. The most preferred animal is the mouse.

Cells that can be used for such inoculations, or for any of the other means used in the invention, include any cell line which naturally expresses a mammalian Y5, or any cell or cell line that expresses a mammalian Y5 or any epitope thereof as a result of molecular or genetic engineering, or that has been treated to increase the expression of a mammalian Y5 by physical, biochemical or genetic means. Preferred cells are human cells, most preferably HEK 293 and BHK cells that have been transformed with a recombinant expression construct comprising a nucleic acid encoding a mammalian Y5, preferably a rat or human Y5, and that express the mammalian Y5 gene product.

The present invention provides monoclonal antibodies that are immunologically reactive with an epitope of mammalian Y5 or fragment thereof and that is present on the surface of mammalian cells, preferably human or mouse cells. These antibodies are made using methods and techniques well known to those of skill in the art.

Monoclonal antibodies provided by the present invention are produced by hybridoma cell lines, that are also provided by the invention and that are made by methods well known in the art. Hybridoma cell lines are made by fusing individual cells of a myeloma cell line with spleen cells derived from animals immunized with cells expressing the Y5 receptor, preferably rat or human cells, as described above. The myeloma cell lines used in the invention include lines derived from myelomas of mice, rats, hamsters, primates and humans. Preferred myeloma cell lines are from mouse. The animals from whom spleens are obtained after immunization are rats, mice and hamsters, preferably mice, most preferably Balb/c mice. Spleen cells and myeloma cells are fused using a number of methods well known in the art, including but not limited to incubation with inactivated Sendai virus and incubation in the presence of polyethylene glycol (PEG). The most preferred method for cell fusion is incubation in the presence of a solution of 45% (w/v) PEG-1450. Monoclonal antibodies produced by hybridoma cell lines can be harvested from cell culture supernatant fluids from in vitro cell growth; alternatively, hybridoma cells can be injected subcutaneously and/or into the peritoneal cavity of an animal, most preferably a mouse, and the monoclonal antibodies obtained from blood and/or ascites fluid.

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Monoclonal antibodies provided by the present invention are also produced by recombinant genetic methods well known to those of skill in the art, and the present invention encompasses antibodies made by such methods that are immunologically reactive with an epitope of a mammalian Y5.

The present invention encompasses fragments of the antibody that are immunologically reactive with an epitope of a mammalian Y5. Such fragments are produced by any number of methods, including but not limited to proteolytic cleavage, chemical synthesis or preparation of such fragments by means of genetic engineering technology. The present invention also encompasses single-chain antibodies that are immunologically reactive with an epitope of a mammalian Y5 made by methods known to those of skill in the art.

The present invention also encompasses an epitope of a mammalian Y5 that is comprised of sequences and/or a conformation of sequences present in the mammalian Y5 molecule. This epitope may be naturally occurring, or may be the result of proteolytic cleavage of the mammalian Y5 molecule and isolation of an epitope-containing peptide or may be obtained by synthesis of an epitope-containing peptide using methods well known to those skilled in the art. The present invention also encompasses epitope peptides produced as a result

of genetic engineering technology and synthesized by genetically engineered prokaryotic or eukaryotic cells.

The invention also includes chimeric antibodies, comprised of light chain and heavy chain peptides immunologically reactive to an epitope that is a mammalian Y5. The chimeric antibodies embodied in the present invention include those that are derived from naturally occurring antibodies as well as chimeric antibodies made by means of genetic engineering technology well known to those of skill in the art.

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Also provided by the present invention are non-human transgenic animals grown from germ cells transformed with the Y5 nucleic acid sequence according to the invention and that express the Y5 receptor according to the invention and offspring and descendants thereof. Also provided are transgenic non-human mammals comprising a homologous recombination knockout of the native Y5 receptor, as well as transgenic non-human mammals grown from germ cells transformed with nucleic acid antisense to the Y5 nucleic acid of the invention and offspring and descendants thereof. Further included as part of the present invention are transgenic animals which the native Y5 receptor has been replaced with the human homolog. Of course, offspring and descendants of all of the foregoing transgenic animals are also encompassed by the invention.

Transgenic animals according to the invention can be made using well known techniques with the nucleic acids disclosed herein. E.g., Leder et al., U.S. Patent Nos.4,736,866 and 5,175,383; Hogan et al., Manipulating the Mouse Embryo, A Laboratory Manual (Cold Spring Harbor Laboratory (1986)); Capecchi, Science 244, 1288 (1989); Zimmer and Gruss, Nature 338, 150 (1989); Kuhn et al., Science 269, 1427 (1995); Katsuki et al., Science 241, 593 (1988); Hasty et al., Nature 350, 243 (1991); Stacey et al., Mol. Cell Biol. 14, 1009 (1994); Hanks et al., Science 269, 679 (1995); and Marx, Science 269, 636 (1995). Such transgenic animals are useful for screening for and determining the physiological effects of Y5 receptor agonists and antagonist. Consequently, such transgenic animals are useful for developing drugs to regulate physiological activities in which NPY and/or PYY participate.

The following Examples are provided for illustrative purposes only and are not intended, nor should they be construed, as limiting the invention in any manner.

EXAMPLES

Example 1

Isolation and Sequencing of Rat Y5 Receptor

Isolation of rat hypothalamus mRNA and construction of cDNA library

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Expression cloning strategy was used to clone novel NPY receptor in rat hypothalamus cDNA library. RNA was obtained from 9 frozen rat hypothalami weighing a total of 0.87 grams. Poly(A) RNA was isolated directly from the tissue using the Promega PolyATtract System 1000 kit (Promega, Madison, WI). The hypothalami were homogenized in 4 mL of 4M guanidine thiocyanate-25mM sodium citrate, pH 7.1-2% B-mercaptoethanol using a Polytron at full-speed for approximately 1 minute. To the homogenized tissue 8 mL of 4M guanidine thiocyanate-25mM sodium citrate, pH 7.1-1% B-mercaptoethanol which had been preheated to 70°C was added. After mixing thoroughly, 870 pmol biotinylated oligo(dT) was added; the mixture was incubated at 70°C for 5 minutes. The homogenate was subjected to centrifugation at 12000 x g for 10 minutes at room temperature; the homogenate was transferred to a clean tube and 10.44 mL Streptavidin MAGNESPHERE® Paramagnetic Particles (SA-PMPs) which had been prepared as per the published protocol was added. (Promega Corp. published protocol TM 228; Promega Corporation, Madison, WI). The homogenate and SA-PMPs were incubated together for 2 minutes at room temperature after which the homogenate was decanted while the SA-PMP-biotinylated oligo(dT)-hypothalamic poly(A) RNA complex was retained in the tube by a magnetic stand. The complex was washed as per the protocol, after which the RNA was precipitated and resuspended in water. 25 micrograms of this poly(A) RNA was used by Invitrogen (Invitrogen Corporation, San Diego, CA) to prepare a cDNA expression library. The protocols used by Invitrogen to prepare the cDNA library are essentially based upon the procedures of Okayama and Berg (Molec. Cell. Biol. 2, 161 (1982)) and Gubler and Hoffman (Gene 25, 263 (1983)) (Invitrogen Corporation publications 130813sa and 130928sa). An oligo(dT) anchor primer was used for reverse transcription, and the library was cloned unidirectionally into pcDNA3 vector which contains a CMV promoter for eukaryotic expression. The cDNA library had 5.3 x 10⁵ primary recombinants with an average insert size of 2.59 kb.

Isolation of a novel Y5 receptor cDNA clone

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The rat hypothalamus cDNA library was plated on the LB/Ampicillin plates in pools of 1,000 independent colonies. The plates were incubated at 37°C for about 20 hours and the bacteria from each plate were scraped in 4-5 ml LB/Ampicillin media. Two ml of the bacteria samples were used for plasmid preparation and one ml of each pool was stored at -80°C in 15% glycerol.

COS-7 cells were grown in Dulbecco's Modified Eagle Medium (DMEM, GIBCO 11965-092), 10% fetal bovine serum (GIBCO 16000-028), and 1 x antibiotic/antimycotic solution (GIBCO 15240-039) (Gaithersburg, MD). Cells were trypsinized and split at 50 to 70% confluency.

DNA from 1300 pools was transfected into COS-7 cells for [125]PYY binding assays. Twenty four hours before transfection, cells were plated into flaskette chambers (Nunc, Inc. 177453, Naperville, IL) at 3×10^5 cells/flaskette (equivalent to 3×10^4 cells/cm²). Two µg of plasmid DNA from each pool was transfected into the cells using 10 µl of Lipofectamine (GIBCO 18324-012) according to the manufacture's protocol. Forty eight hours after transfection, the [125I]PYY binding assay was performed in the flaskette chamber. The cells were treated with 2 ml total binding buffer: 10 mM HEPES, 5 mM KCl, 1.2 mM KH₂PO₄, 2.5 mM CaCl₂, 1.2 mM MgSO₄, 150 mM NaCl, 25 mM NaHCO₃, 10 mg/ml bovine serum albumin, 0.5 mg/ml bacitracin and 0.4 mg/ml soybean trypsin inhibitor at room temperature for 15 minutes. The cells were then incubated with 100 pM porcine [125I]PYY (Amersham (Arlington Heights, IL), Specific Activity 4000Ci/mmol) in the total binding buffer for 90 minutes at room temperature. After binding, the cells were washed three times with ice-cold total binding buffer without ligand and one time with cold phosphate buffered saline (PBS). Cells were fixed with 1% cold glutaraldehyde in PBS for 15 minutes, washed once with cold PBS/0.5 M Tris, pH 7.5 and incubated in PBS/0.5 M Tris, pH 7.5 for 15 minutes at 4°C. After washing one more time with cold PBS, the slides were dipped in 0.5% gelatin at 42°C and dried The dried slides were dipped in 50% photographic emulsion (Kodak under vacuum. (Rochester, NY) NTB2) at 42°C and exposed in the darkbox for four days at 4°C. After four days of exposure, the darkbox was moved to room temperature for one hour and slides were developed in developer D-19 (Kodak) for three minutes at 15°C and fixed in fixer (Kodak) for three minutes at 15°C, washed in water and air dried. Cells were stained with Diff-Quik stain

set (Baxter, McGaw Park, IL) and air dried. Slides were dipped into xylenes and mounted with DPX mountant (Electron Microscopy Science, Fort Washington, PA). Positive cells were identified using dark field microscopy.

Twenty one positive pools were identified. Since the hypothalamus expresses different subtypes of NPY receptors including Y1 and Y2 receptors, we analyzed all the positive pools for Y1, Y2 and Y4/PP receptors by PCR. Of the 21 positive pools tested as described above, 12 pools contained Y1, 4 pools contained Y2 and none contained Y4/PP. Five pools (Y217, Y555, Y589, Y861 and Y1139) were negative by PCR analysis. The pool Y217 was subdivided in 24 subpools of 200 colonies, then 50 colonies, and finally a single clone, the Y217.24.13.6B clone (6B), was isolated.

DNA and peptide sequences analysis

Plasmid DNA was sequenced by Lark Technologies Inc. (Houston, Texas) and Biotechnology Resource Laboratory of Yale University (New Haven, CT) using Sequenase Kit (US Biochemical, Cleveland, OH) or Applied Biosystems' automatic sequencer system (model 373A). The peptide sequence was deduced from the long open-reading-frame of the nucleotide sequence. DNA and peptide sequences were analyzed using the GCG program (Genetics Computer Group, Madison, WI). The results are embodied in SEQ ID NO 1 (the nucleic acid sequence) and SEQ ID NO 2 (the amino acid sequence).

Example 2

Localization of Rat Y5 Receptor in Brain and Other Tissues

Northern Blot

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To study the expression level of the Y5 receptor in the rat brain and other tissues, we did Northern blot analysis using the 6B 2.4 kb probe. A rat multiple tissue Northern blot (Clontech Laboratories, Palo Alto, CA) was hybridized to the ³²P-labeled rat 6B probe. The blot contains 2 µg of poly A⁺ RNA per lane from rat heart, brain, spleen, lung, liver, skeletal muscle, kidney, and testis. Hybridization was carried out in 1x hybridization solution containing 6X SSC (0.9 M NaCl, 0.09 M Na Citrate, pH 7.0), 5x Denhardt's solution (0.1% polyvinylpyrrolidone, 0.1% ficoll type 400, 0.1% bovine serum albumin), 100 mg/ml sheared, and denatured salmon sperm DNA at 65°C. The filter was washed at 65°C in 0.1X SSC, 0.1% SDS and exposed to Kodak XAR 5 film with two intensifying screens. A single 2.6 kb band was detected in the brain after

overnight exposure of the blot. No bands were found from other tissues (heart, spleen, lung, liver, skeletal muscle, kidney and testis) in the Clontech multiple tissue Northern blot, even after six days of exposure.

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We tested 6B expression in more rat tissues and different regions of brain. mRNA was isolated from rat whole brain, cortex, hypothalamus, hippocampus, olfactory bulb, spleen, stomach, kidney, small intestine, adrenal and pancreas using Fast Track Isolation Kit (Invitrogen). Ten µg of mRNA from different brain regions and multiple tissues were run on a denaturing formaldehyde 1% agarose gel, transferred to a Nytran membrane (Schleicher and Schuell) and hybridized with ³²P-labeled 6B 2.4 kb probe and washed at high stringency. After overnight hybridization, the filter was washed at high stringency and exposed to X-ray film with intensifying screens. The 6B receptor mRNA was detectable in the brain regions examined after one day exposure, but no signal was observed from other tissues, even after a week exposure with double intensifying screens.

Example 3

Isolation of Two Isoforms of the Rat Y5 Receptor

Plasmid DNA from pools Y555, Y589, and Y861 described in Example 1 were hybridized to the Y5 probe at high stringency. A single positive clone was isolated from the Y555 pool and sequenced as described in Example 1. Compared to the 6B DNA sequence, the Y555 sequence (SEQ ID NO 4 has a 123 bp insert sequence located at the 5'-untranslated region between nucleotides 239 and 240 of Y5 clone. The coding region of the clones Y555, Y589, and Y861 has the same sequence as clone 6B, except for one nucleotide substitution (C to T) at position 430 of the 6B clone. The nucleotide substitution changes the amino acid proline to leucine in the first transmembrane domain. The corresponding amino acid sequence is given by SEQ ID NO 4.

The different isoforms of the receptor could be the allelic variants of the same gene. To test this hypothesis, we analyzed genomic DNA from 16 rats. The genomic DNA from each animal was used as template for PCR analysis. A 314 bp DNA fragment that contains the site of the nucleotide variation was amplified and sequenced. Of the 16 DNA samples tested, 14 samples had a T at position 430 and 2 samples had a C. This result strongly suggests that the amino acid variation is an allelic variant.

Example 4

Pharmacological Characterization of the Novel Rat NPY Receptors

Transient Transfection

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Monkey kidney cells (COS-7) were maintained in T-175 cm² flasks (NUNC) at 37°C with 5% CO₂ in a humidified atmosphere. Cells were grown in Dulbecco's Modified Eagle Medium (DMEM) supplemented with 2 mM glutamine, 10% fetal bovine serum, 1 mM sodium pyruvate, and antibiotic/antimycotic. Cells at 70% confluency were transfected with Y5 DNA using the Lipofectamine method (GIBCO-BRL). 15 µg DNA and 90 µl Lipofectamine were added to each flask. Media was completely replaced 24 hours post transfection, and membranes were harvested 24 hours later.

Stable Expression of the Rat NPY Y5 receptor (clone Y861)

A strain of the human embryonic kidney cell line 293 adapted to grow in suspension (293S) was used for these experiments. Approximately 1x10⁶ cells were seeded onto a 10-cm dish 24 hours prior to transfection. The rat NPY Y5 cDNA (Y861), subcloned in the eukaryotic expression vector pcDNA3 (Invitrogen, Carlsbad, CA) was first linearized with Notl and purified using a Wizard PCR Prep kit (Promega). In preparation for transfection, 15 µg of the linearized DNA were added to 500 µl of DMEM cell culture media, and 30 µl of Lipofectamine (Life Sciences) were added into a separate 500 µl aliquot of DMEM. These two solutions were mixed together and incubated for 20 minutes at room temperature and the resulting DNA/lipid complexes were then slowly added to the cells (which had been previously rinsed once with serum-free DMEM) and covered with a total volume of 10 ml. Cells were then transferred to a humidified 10% CO₂ incubator and left for 4 hours at 37°C, at which time the media was replaced with DMEM supplemented with 8% FBS. After 16 hours, cells were trypsinized and split at a 1:15 ratio into 10-cm dishes containing DMEM/8% FBS in the presence of 700 µg/ml of G418 (selection media). When discrete colonies became apparent (after approximately 10 days), cells were pooled and carried through 2 additional passages in selection media. Cells were then trypsinized and diluted in preparation for cloning by limited dilution (CBLD), such that an average of one cell was seeded in each well of a 96-well microtiter culture plate, and was inspected periodically for the subsequent 2 to 3 weeks. After 21 days in culture under selection conditions, those wells containing single colonies were selected and transferred to 24well culture plates following trypsinization. Each of these clones was propagated until

sufficient quantities were available for testing [125I]PYY binding activity, from which one particular clone designated E7 was selected on the basis of its high level of binding activity.

Stable Expression of the human NPY Y5 receptor

293 cells were plated onto a T75 flask one day prior to transfection such that they were 50-70% confluent when used for the experiment. The human NPY Y5 intronless genomic clone HG.PCR15, containing the full length open reading frame encoding the receptor, was first linearized with *Not* I and purified using a Wizard PCR Prep kit (Promega). For each transfection, 8 μg of linearized DNA were added to 1.25 ml of Optimem culture media (Life Sciences) and 37 μl of Transfectam (Promega) were added to 1.25 ml of Optimem. These two solutions were then mixed together and added to cells previously washed once with Optimem. After an incubation period of 5 hours, the DNA/Transfectam mixture was removed, cells were washed with PBS and fed with DMEM supplemented with 10% FBS. Cells were left intact for two days, and then switched to selection media (DMEM 10% FBS containing 350 μg/ml of G418) for 5-10 days followed by CBLD as described above. The individual clone 293.hy5.sb.8 was selected on the basis of its high level of [125]PYY binding activity, using the intact cell binding protocol from above.

Membrane Preparation

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The media was removed from each flask of transfected cells, and the cells were washed twice with 20 ml ice-cold phosphate buffered saline. The cells were scraped from the flask in 5 ml of Tris buffer (20 mM Tris-HCl and 5 mM EDTA, pH 7.7), and then transferred to a centrifuge tube. Each flask was washed with an additional 5 ml of Tris buffer and combined in the centrifuge tube. The cells were polytroned for 2 x 10 seconds (12 mm probe, 7000-8000 rpm) and centrifuged 5 minutes (Centra 7R, International Equipment Co., Needham Heights, MA) at 800 rpm and 4°C. The supernatant was then transferred to a clean centrifuge tube and was centrifuged at 30,000 x g for 30 minutes and 4°C. The supernatant was removed and the pellet was stored at -80°C. Protein concentration was measured using the Bio-Rad kit pursuant to the standard manufacturer's protocol (Biorad Laboratories, Hercules, CA) with bovine IgG as the standard.

sufficient quantities were available for testing [125I]PYY binding activity, from which one particular clone designated E7 was selected on the basis of its high level of binding activity.

Stable Expression of the human NPY Y5 receptor

293 cells were plated onto a T75 flask one day prior to transfection such that they were 50-70% confluent when used for the experiment. The human NPY Y5 intronless genomic clone HG.PCR15, containing the full length open reading frame encoding the receptor, was first linearized with *Not* I and purified using a Wizard PCR Prep kit (Promega). For each transfection, 8 μg of linearized DNA were added to 1.25 ml of Optimem culture media (Life Sciences) and 37 μl of Transfectam (Promega) were added to 1.25 ml of Optimem. These two solutions were then mixed together and added to cells previously washed once with Optimem. After an incubation period of 5 hours, the DNA/Transfectam mixture was removed, cells were washed with PBS and fed with DMEM supplemented with 10% FBS. Cells were left intact for two days, and then switched to selection media (DMEM 10% FBS containing 350 μg/ml of G418) for 5-10 days followed by CBLD as described above. The individual clone 293.hy5.sb.8 was selected on the basis of its high level of [125]PYY binding activity, using the intact cell binding protocol from above.

Membrane Preparation

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The media was removed from each flask of transfected cells, and the cells were washed twice with 20 ml ice-cold phosphate buffered saline. The cells were scraped from the flask in 5 ml of Tris buffer (20 mM Tris-HCl and 5 mM EDTA, pH 7.7), and then transferred to a centrifuge tube. Each flask was washed with an additional 5 ml of Tris buffer and combined in the centrifuge tube. The cells were polytroned for 2 x 10 seconds (12 mm probe, 7000-8000 rpm) and centrifuged 5 minutes (Centra 7R, International Equipment Co., Needham Heights, MA) at 800 rpm and 4°C. The supernatant was then transferred to a clean centrifuge tube and was centrifuged at 30,000 x g for 30 minutes and 4°C. The supernatant was removed and the pellet was stored at -80°C. Protein concentration was measured using the Bio-Rad kit pursuant to the standard manufacturer's protocol (Biorad Laboratories, Hercules, CA) with bovine IgG as the standard.

[125] UPYY Binding Assay for NPY Y5 Receptors

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The binding assays were performed on GF/C Millipore (Bedford, MA) 96-well plates pretreated with 0.02% polyethylenimine (PEI) for at least 2 hours prior to use. The PEI was aspirated from the plates on a vacuum manifold immediately before the samples were added to the wells. All peptides, tissue and radioligand were diluted with binding buffer (25 mM Tris, 120 mM NaCl, 5 mM KCl, 1.2 mM KH₂PO₄, 2.5 mM CaCl₂, 1.2 mM MgSO₄, 0.1% BSA and 0.5 mg/ml bacitracin, pH 7.4). For competition assays, increasing concentrations of peptide were incubated with [¹²⁵I]PYY and tissue. In a final volume of 200 µl, samples consisted of: membrane protein (*i.e.*, 2.5-15 or 10-30 µg membrane protein for rat Y5 or human Y5, respectively); 75-100 pM [¹²⁵I]PYY NEN-DuPont (Boston, MA); peptide dilution or binding buffer. Nonspecific binding was defined by 1 µM PYY. NPY, PYY, (2-36)NPY, (10-36)NPY, (LP)(3-36)NPY and (32D-Trp)NPY were synthesized at Bayer Corp. (West Haven, CT). All other peptides were purchased from either Peninsula (Belmont, CA) or Bachem (Torrance, CA).

For saturation experiments, increasing concentrations of [125]PYY were incubated with membrane and 1 µM PYY. After a 2 hour incubation at room temperature with constant mixing, the samples were aspirated on a vacuum manifold. The wells were washed with three 200 µl aliquots of ice-cold binding buffer. The individual wells were punched into 12x75 mm plastic tubes, and counted on a Wallac (Gaithersburg, MD) gamma counter. Binding data were analyzed using the nonlinear regression curve-fitting program RS/1 (BBN Software Products Corp., Cambridge, MA).

Binding Assays for Rat Y2, Y1, and Y4/PP1 Receptors

The binding buffer for rat Y2 binding was Krebs/Ringer bicarbonate (Sigma K-4002, S-8875), pH 7.4, containing 0.01% bovine serum albumin (BSA - Sigma A-2153) and 0.005% bacitracin. 0.85-1 μ g of protein and 25 pM [¹²⁵I]PYY are added to each well. Nonspecific binding is defined by 1 μ M NPY.

The binding buffer for rat Y1 and rat Y4/PP1 binding consisted of 137 mM NaCl, 5.4 mM KCl, 0.44 mM KH₂PO₄, 1.26 mM CaCl₂, 0.81 mM MgSO₄, 20 mM HEPES, 1 mM dithiothreitol (DTT), 0.1% bacitracin, 100 mg/l streptomycin sulfate, 1 mg/l aprotinin, 10 mg/ml soybean trypsin inhibitor and 0.3% BSA, pH 7.4. For rat Y1 binding, ~5-15 μg of protein and 50 pM [¹²⁵I]PYY were added to each well, and nonspecific binding was defined by 1 μM NPY. For the rat Y4/PP1 binding assay, ~1-2 μg of protein and 50 pM rat [¹²⁵I]PP (NEN

DuPont, Boston, MA) were added to each well, and 1 μ M rat PP was used to define nonspecific binding.

In Vitro Functional Assay - Measurement of Forskolin-Stimulated Adenylate Cyclase

Rat Y5

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(Reference: Gordon et al., J. Neurochem. 55, 506, 1990) Suspension cells stably expressing the Y5 receptor (approximately 400,000 per sample) were resuspended in serum-free DMEM containing 10mM HEPES (pH 7.4) and 1 mM isobutylmethylxanthine (IBMX). 1uM forskolin was added to the cells. The assay was stopped by transferring the samples into a boiling water bath for 3 minutes. After a 3 minute centrifugation at 14,000xg, an aliquot of each sample was quantitated for cAMP levels by radioimmunoassay (NEN DuPont, MA).

Human Y5

Monolayer cells stably expressing the Y5 receptor were pre-rinsed with Wash buffer (pH 7.2: 137 mM NaCl, 2.7 mM KCl, 0.9 mM CaCl₂, 0.5 mM MgCl₂, 6.5 mM Na₂HPO₄, 1.5 mM KH₂PO₄). Cells were then incubated for 10 minutes at 37°C in Assay buffer (pH 7.4: Wash buffer + 10 mM HEPES, 10 mg/ml BSA, 0.5 mg/ml bacitracin, 0.4 mg/ml soybean trypsin inhibitor). After addition of fresh buffer and 100 μM IBMX, the cells were incubated for 10 minutes at 37°C. The reaction was started with the addition of peptide and 1-10 μM forskolin. After a 20 minute incubation at 37°C, the reaction was terminated by discarding the buffer and adding 65% ethanol to each well. The supernatant was then transferred to microfuge tubes and the extraction step was repeated once more. After evaporation of the ethanol from the samples, the amount of cAMP was assayed using by radioimmunoassay (NEN DuPont: Boston, MA).

In Vivo Pharmacology Procedures

Adult male Wistar rats were surgically implanted with a chronic intracerebral ventricular (ICV) cannula (Plastic Products, Roanoke, Virginia) using a stereotaxic instrument. Several days after the surgery, 1-6 nmoles of each peptide (or saline) was injected into the lateral ventricle of 4-12 rats in a volume of 5-10 µl. The quantity of rodent chow consumed in a 2 hour period was measured.

In Vitro and In Vivo Pharmacology Results

Figure 1 presents the competition curves of various peptides for [125 I]PYY binding to Y5 receptor membranes transiently expressed in COS-7 cells. Each point is the average value of triplicate determinations from a representative experiment. IC₅₀ values corresponding to 50% inhibition of specific binding were determined using nonlinear regression analysis. K_i values were calculated from the IC₅₀ values using the Cheng-Prusoff correction, such that $K_i = IC_{50}/(1 \pm (L/K_d))$, where L is the radioligand concentration and K_d is the dissociation constant. The results for transiently expressed Y5 clones are presented in Table 1, and Table 2 contains data for stably expressed Y5 clones.

Table 1

PEPTIDE	NPY Y5 BINDING AFFINITIES (K, ± SEM, nM)												
	RAT Y555 Clone*	RAT 6B Clone	RAT Y861 Clone	HUMAN Clone									
r/hNPY	0.53 ± 0.06	0.49 ± 0.03	0.50 ± 0.06	0.73 ± 0.09									
rPYY	1.1 (1.2, 0.95)	0.48 ± 0.09	1.0 ± 0.13	1.3 ± 0.14									
h(LP)PYY	2.5 ± 0.5	0.57 ± 0.01	1.8 ± 0.09	1.7 ± 0.3									
r/h(LP)NPY	0.96 (1.0, 0.92)	0.31	0.55 ± 0.11	0.97 ± 0.36									
p(LP)NPY	ND	0.64 ± 0.07	0.47	0.88 ± 0.11									
r/h(2-36)NPY	0.81 (0.61, 1)	0.65	1.2 ± 0.07	1.2 ± 0.15									
p(3-36)NPY	3.6 ± 0.4	1.9 ± 0.27	2.0 (1.8, 2.3)	10.4 ± 2.0									
r/h(3-36)NPY	ND	0.49	2.1 (2.7, 1.6)	3.8 <u>+</u> 0.48									
r(3-36)PYY	6.2 ± 1.1	1.4 ± 0.10	4.2 ± 0.47	10 ± 3.4									
r/h(10-36)NPY	35	4.9 (6.0, 3.8)	34 ± 2.8	110 (110, 109)									
p(13-36)NPY	40 (38, 41)	7.7 (7.9, 7.5)	22 (25, 19)	56 ± 7									
r(13-36)NPY	73	11 ± 1.0	86 ± 19	77 (89, 65)									
p(18-36)NPY	303	194 ± 88	206 ± 61	618 ± 85									
r/h(20-36)NPY	636	330 ± 31	587	>1000									
r/h(22-36)NPY	>1000	>1000	>1000	>1000									
r/h(26-36)NPY	>1000	>1000	>1000	>1000									
(1-24)NPY	ND	>1000	>1000	>1000									
BIBP3226	ND	>1000	>1000	>1000									
hPP	ND	ND	4.0 ± 0.29	11 (15, 6.2)									
rPP	ND	62	296 ± 47	436 (582, 290)									

^{*} IC50 values (nM)

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Table 2

Peptide	K, Values (nM; Average ± SEM)									
	293.hY5.sb.8	293S.Y861.2								
rPYY	1.3 ± 0.2	0.71 ± 0.1								
hPYY	1.1 ± 0.2	1.06 ± 0.2								
(3-36)PYY	4.5 ± 0.7	3.6 ± 0.4								
(13-36)PYY	24 ± 2.0	29 ± 4								
h(LP)PYY	1.3 ± 0.1	0.76 ± 0.1								
r/hNPY	0.79 ± 0.1	0.86 ± 0.07								
p(LP)NPY	1.2 ± 0.4	0.67 ± 0.04								
h/r(LP)NPY	0.89 ± 0.1	0.67 ± 0.04								
(LP)(3-36)NPY	3.1 ± 0.6	2.9 ± 0.9								
(2-36)NPY	1.4 ± 0.03	0.83 ± 0.1								
h(3-36)NPY	3.5 ± 0.4	1.4 ± 0.4								
(10-36)NPY	14 ± 2.7	15 ± 4.7								
p(13-36)NPY	8.7 ± 1.6	8.8 ± 2.0								
p(18-36)NPY	144 ± 18	61 ± 13								
(20-36)NPY	429 ± 133	108 ± 16								
(22-36)NPY	>900	>930								
(26-36)NPY	>900	>930								
(1-24)NPY	>900	>930								
(32D-Trp)NPY	7.3 ± 0.8	4.2 ± 1.0								
hPP	3.7 ± 1.6	2.5 ± 0.5								
rPP	286 ± 77	203 ± 44								

 K_1 values for various peptides for [^{125}I]PYY binding to the transiently expressed rat 6B, Y861 and Y555 receptor clones as well as the human Y5 receptor. The averages \pm standard error of the mean (SEM) represent values from at least three independent experiments. Two independent experiments are represented by the average, followed by the individual values in parentheses. Remaining values without SEM are from a single experiment. Peptide species in Table 1 (and Table 2, *infra*) are indicated with the following prefixes: r = rat, h = human, p = porcine, r/h = rat = human. ND = not determined.

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The rank order of the affinities of the peptides tested is as follows:

$$NPY \sim PYY \sim (LP)PYY \sim (LP)NPY \sim (2-36)NPY \sim (3-36)PYY \sim$$
 $(LP)(3-36)NPY \sim (3-36)NPY > (32D-Trp)NPY > (10-36)NPY \sim (13-36)NPY >$
 $(18-36)NPY > (20-36)NPY > (22-36)NPY, (26-36)NPY$

In Table 3, the pharmacological profile of the standard peptides is expanded for the other cloned NPY receptors to further illustrate the novel nature of the Y5 receptor pharmacology. In addition, the *in vivo* feeding response of some of these peptides is listed for comparison. The data shown are representative of the average of at least two independent experiments, as described in the methods. Feeding of rats injected (ICV) with saline was < 3g/2hours.

Table 4 shows the EC₅₀ values for same standard peptides at the rat and human Y5 receptor.

C-terminal fragment (3-36)NPY binds preferentially to Y2 receptors, while (LP)NPY has lower affinity. Conversely, (LP)NPY has high affinity for the Y1 receptor, while (3-36)NPY and the C-terminal fragments are much weaker. When considering the rat Y4/PP1 receptor, rat PP has very high affinity as compared to NPY, PYY, (LP)NPY, and (13-36)NPY. In the *in vivo* feeding model, (LP)NPY, which has high affinity for Y1 and low affinity for Y2, and (3-36)NPY, which has a high affinity for Y2, but not Y1, all stimulate feeding in rats. Rat PP does not induce much feeding when administered to rats. This *in vivo* profile matches the *in vitro* pharmacological profile outlined in Table 2 for the Y5 receptor.

In addition, while (LP)(3-36)NPY (a custom peptide synthesized at Bayer) has weak affinity for Y1, Y2 and Y4/PP1, it stimulates feeding in rats. Importantly, (LP)(3-36)NPY has high affinity for the Y5 receptor (Table 2). These data are further evidence that the Y5 receptor is linked to feeding.

Table 3

IC ₅₀ VALUE (nM)												
PEPTIDE	Rat Y1 (clone)	Rat Y2 (clone)	Rat Y4/PP1 (clone)	Rat Y5 (Y861)	Feeding (g/2 h)							
r/hNPY	0.13	0.24	> 1000	0.45	>5							
rPYY	0.43	0.079	630	0.9	>5							
h(Leu ³¹ Pro ³⁴)PYY	0.57	116	ND	2.0	>5							

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IC ₅₀ VALUE (nM)													
PEPTIDE	Rat Y1 (clone)	Rat Y2 (clone)	Rat Y4/PP1 (clone)	Rat Y5 (Y861)	Feeding (g/2 h)								
p(Leu ³¹ Pro ³⁴⁾ NPY	0.15	150	4.3	0.63	>5								
r/h(2-36)NPY	47	0.50	>1000	1.3	>5								
p(3-36)NPY	45	0.67	>1000	2.2	>5								
r/h(Leu ³¹ Pro ³⁴) (3-36)NPY	44	154	20	3.4	>5								
hPP	40	>1000	0.065	4.9	>5								
(32DTrp)NPY	> 1000	26	ND	7.0	ND								
r/h(10-36)NPY	148	0.42	>1000	34	<3								
rPP	843	>1000	0.071	325	- <3								
p(18-36)NPY	287	0.34	159	326	<3								
(20-36)NPY	435	0.64	ND	638	<3								
(22-36)NPY	(22-36)NPY >1000		ND	>1000	<3								
(26-36)NPY	>1000	84	ND	>1000	<3								
(1-24)NPY	>1000	>1000	ND	>1000	<3								

The pharmacological profile for the 6B (and Y861 and Y555) receptor clones is distinct from Y1 receptors (where PYY~NPY~(LP)NPY > (3-36)NPY > (13-36)NPY ~ (18-36)NPY > (LP)(3-36)NPY), as well as Y2 receptors (where PYY~NPY~(13-36)NPY~(18-36)NPY~(3-36)NPY >> (LP)NPY~(LP)(3-36)NPY). The Y5 receptor is also different from the pancreatic polypeptide (PP) receptor (Y4/PP) since [1251] PP (rat) does not bind to it.

Although the rank order of affinities is essentially the same when comparing 6B to Y861 and Y555, subtle differences do exist in the IC₅₀ values. It appears that Y861 and Y555 have slightly lower affinities (approximately 2- to 3-fold) for PYY and other PYY analogs, as compared to 6B. In addition, (10-36)NPY and (13-36) have 2- to 4-fold lower affinity for Y861 and Y555.

Nonlinear regression analysis of saturation data for the Y5 receptor yielded a K_d value of 0.27 nM and a receptor density (B_{max}) of about 140 fmol/mg protein in these transiently transfected cells.

Fig. 2 presents the saturation curve for specific binding of [^{125}I]PYY to Y5 receptor membranes transiently expressed in COS-7 cells. Membranes were incubated with concentrations of [^{125}I]PYY ranging from 0.05 to 5 nM, in the presence or absence of 1 μ M PYY. Each point represents the average value of triplicate determinations at each concentration tested. The inset in Fig. 2 shows the corresponding Rosenthal plot of the data.

Peptide	EC ₅₀ Values, nM (n Value)								
	293S.Y861.2	293.hY5.sb.8							
r/hNPY	6.3 ± 1.9 (3)	0.3 (1)							
rPYY	6.5 (2)	ND							
r/h(2-36)NPY	21 (2)	ND							
r/h(3-36)NPY	ND	6(1)							
r/h(LeuPro)(3-36)NPY	31 ± 39 (3)	23 ± 11 (3)							
(32D-Trp)NPY	24 (1)	33 (1)							
hPP	1 (1)	5 (1)							
rPP	. 112(1)	>1000 (1)							

Table 4

Example 5

Isolation of Human Y5 Receptor

10 Isolation of Human Genomic Clone

Polymerase chain reaction (PCR) was used to amplify a 375 base pair (bp) coding region of the rat Y5 cDNA clone. The primers for the PCR were:

- (+) 5'-TAGGGAACCTGGCCTCCTCC-3' (SEQ ID NO 5) (nucleotides 487-506),
- (-) 5'-TCAGAGGGCCATGACTCAAC-3' (SEQ ID NO 6) (nucleotides 843-862).
- The PCR product was cloned into pCRII vector (Invitrogen) and sequenced. After confirmation by sequencing, the insert was purified from the low melting gel and labeled with digoxigenin-11-dUTP using the random primed method (Boehringer Mannheim, Indianapolis, IN). The labeled probe was used to screen human genomic library.

1x10⁶ independent recombinants were screened from the library. Filter hybridization was carried out in the hybridization buffer containing 6x SSC, 0.1% N-lauroylsarcosine, 0.02% sodium dodecyl sulfate (SDS), 3% blocking reagent (Boehringer Mannheim) and 30% formamide at 37°C overnight. The filters were washed at 37°C in 0.1x SSC, 0.1% SDS and the positive clones were identified by CSPD detection kit according to the manufacturer's protocol (Boehringer Mannheim).

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Two positive clones (HG11A and HG19) were isolated from the library. The positive clones were subcloned into pBluescript vector (Stratagene). One clone, h11a, was analyzed by restriction mapping and plasmid Southern blot. Two EcoRV fragments, 2.4 kb and 0.4 kb, were hybridized by the rat Y5 probe. These two DNA fragments were subcloned and sequenced from both ends. DNA sequence analysis was performed using GCG program. The coding region of the human Y5 genomic clone was identified by DNA sequence analysis. This region was amplified by PCR using genomic clone h11A as template and subcloned into pcDNA3 expression vector (Invitrogen) for further studies. The h11A clone has the nucleic acid coding sequence given by SEQ ID NO 5 and the protein that it encodes has the amino acid sequence given by SEQ ID NO 6.

The human Y5 DNA coding region was used to search the sequence similarities in the gene bank. The Y5 coding sequence from nucleotide 821 to the stop codon at position 1338 is nearly identical, but in an opposite orientation, to part of the human NPY-Y1 gene (Ball et al, J. Biol. Chem. 270, 30102 (1995)). The identical sequence covered the 1C exon promoter, exon 1C, and part of the intron sequences of the NPY-Y1 receptor in an opposite orientation. Compared to the published nucleotide sequence, the Y5 coding region has a T insertion at position 1226 and a TG insertion at positions 1235 and 1236.

SEQUENCE LISTING

5	(1) GENER	RAL INFORMATION:
3	(1)	APPLICANT: Hu Ph.D., Yinghe
	(1)	McCaleb Ph.D., Michael L.
		Bloomquist Ph.D., Brian T.
		Flores-Riveros Ph.D., Jaime R.
10		Cornfield Ph.D., Linda J.
	(ii)	TITLE OF INVENTION: Neuropeptide Y Receptor and Nucleic Acid Sequences
15	(iii)	NUMBER OF SEQUENCES: 8
	(iv)	CORRESPONDENCE ADDRESS:
		(A) ADDRESSEE: McDonnell Boehnen Hulbert & Berghoff
		(B) STREET: 300 South Wacker Drive
20		(C) CITY: Chicago
	•	(D) STATE: IL
		(E) COUNTRY: USA
		(F) ZIP: 60606
25	(37)	COMPUTER READABLE FORM:
		(A) MEDIUM TYPE: Floppy disk
		(B) COMPUTER: IBM PC compatible
		(C) OPERATING SYSTEM: PC-DOS/MS-DOS
		(D) SOFTWARE: PatentIn Release #1.0, Version #1.30
30		
	(vi)	CURRENT APPLICATION DATA:
		(A) APPLICATION NUMBER:
		(B) FILING DATE:
35		(C) CLASSIFICATION:
33	(viii)	ATTORNEY/AGENT INFORMATION:
	(,	(A) NAME: Greenfield Ph.D., Michael S.
		(B) REGISTRATION NUMBER: 37,147
		(C) REFERENCE/DOCKET NUMBER: 96,149/WH 405
40		
	(ix)	TELECOMMUNICATION INFORMATION:
		(A) TELEPHONE: (312)715-1000
		(B) TELEFAX: (312)715-1234
45		
	(2) INFO	PRMATION FOR SEQ ID NO:1:
	(i)	SEQUENCE CHARACTERISTICS:
		(A) LENGTH: 2481 base pairs
50		(B) TYPE: nucleic acid
		(C) STRANDEDNESS: single
		(D) TOPOLOGY: linear
55	(ii)	MOLECULE TYPE: CDNA

(iv) ANTI-SENSE: NO

		(ix)		URE:			DC.										
5				LOC				1589	5								
		(ix)		TURE:		EY: π	at_p	epti	ide						•		
10			(B)	LOC	ATIC	ON: 2	48	1582	2								
		1	220	JENCE			1TT ()	7. C1	en ti	NO.	. 1 .						
																	60
15																ICTCA	60
••	GGGA	.CTGT	CA C	GTGT	יכככי	G AG	STGC	TCT	ĀĀĀ	ACCC'	TGG (CGGC	rccg	GA GC	ccc	TCCTT	120
	CCCA	CCAC	CG C	crcci	AGGT	C CT	CTC	TGC	CGC	CACC	GCT 1	ICCA	TCTG	GA GO	ADA	AGCGA	- 180
20	CCGC	GCTC	AG C	CACG:	racc	c cœ	GAGT	CCAG	GCA	cccg	CAG (CGGC	CGGG	GC A	rccc	GAGGA	240
	TTT	AGT	ATG Met	GAG '	rrr Phe	AAG (CTT (GAG Glu	GAG Glu	CAT His	TTT . Phe .	AAC Asn	AAG :	ACA '	TTT Phe	GTC Val	289
26			1			-	5					10					
25	ACA	GAG	AAC	AAT .	ACA	GCT	GCT	GCT	CGG	AAT	GCA	GCC	TTC	CCT	GCC	TGG	337
	Thr 15	Glu	Asn	Asn '	Thr	Ala 20	Ala .	Ala	Arg	Asn	25	Ala	Pne	PIO .	ALA	Trp 30	
30	GAG	GAC	TAC	AGA	GGC	AGC	GTA	GAC	GAT	TTA	CAA	TAC	TTT	CTG	ATT	GGG	385
	Glu	qaA	Tyr	Arg	Gly 35	Ser	Val	Asp	Asp	Leu 40	Gln	Tyr	Phe	Leu	45	GIY	
	ر با ر	тат	aca	TTC	GTA	TD4	CTT	CTT	GGC	TTT	ATG	GGC	AAT	CTA	CCT	TTA	433
35	Leu	Tyr	Thr	Phe 50	Val	Ser	Leu	Leu	Gly 55	Phe	Met	Gly	Asn	Leu 60	Pro	Ile	
	TTA	ATG	GCT	GTT	ATG	AAA	AAG	CGC	AAT	CAG	AAG	ACT	ACA	GTG	AAC	TIT	481
40	Leu	Met	Ala 65	Val	Met	Lys	Lys	Arg 70	Asn	Gln	Lys	Thr	Thr 75	Val	Asn	Pne	
	CTC	ATA	GGC	AAC	CTG	GCC	TTC	TCC	GAC	ATC	TTG	GTC	GTC	CTG	TTT	TGC	529
	Leu	Ile 80		Asn	Leu	Ala	Phe 85	Ser	qaA	Ile	Leu	90	Val	Leu	Pne	Cys	
45	TCC	· ~~		200	CTG	» CC	ጥርጥ	GTC	TTG	TTG	GAT	CAG	TGG	ATG	TTT	GGC	577
	Ser	Pro	Phe	Thr	Leu	Thr	Ser	Val	Leu	Leu	qeA .	Gln	Trp	Met	Phe	GIY	
	95	i				100			•		105					110	
50	AAA	GCC	ATG	TGC	CAT	ATC	ATG	CCG	TTC	CTI	CAA	TGT	GTG Val	TCA Ser	GTI Val	Leu	625
	rye	s Ala	. Met	. сув	H18		rie C	PLC	, FIIC	120)	,u	- 		125	5	
	GTT	r TCI	A ACT	CTG	ATT	TTA	ATA .	TC	ATI	GCC	TTA :	GTC	agg	TAT	CA	DTA T	673
55	Va]	l Sei	r Thi	Leu 130	Ile	. Leu	Ile	Sei	135	: Ala	ı Ile	e Val	l Arg	140	HI	s Met	

	ATA Ile	AAG Lys	CAC His 145	CCT Pro	ATT Ile	TCT Ser	DAA neA	AAT Asn 150	TTA Leu	ACG Thr	GCA Ala	AAC Asn	CAT His 155	GGC Gly	TAC Tyr	TTC Phe	721
5	Leu	11e 160	Ala	Thr	Val	TGG Trp	Thr 165	Leu	Gly	Phe	Ala	Ile 170	Cys	Ser	Pro	Leu	769
10						CTT Leu 180											817
15						TAT Tyr											865
20						ACA Thr											913
	Pro	Leu	Val 225	Сув	Leu	ACG Thr	Val	Ser 230	His	Thr	Ser	Val	Суз 235	Arg	Ser	Ile	961
25	Ser	Сув 240	Gly	Leu	Ser	CAC	Lув 245	Glu	Asn	Arg	Leu	Glu 250	Glu	Asn	Glu	Met	1009
30	Ile 255	Asn	Leu	Thr	Leu	CAG Gln 260	Pro	Ser	Lys	Lys	Ser 265	Arg	Asn	Gln	Ala	Lys 270	1057
35	Thr	Pro	Ser	Thr	Gln 275	Lys	Trp	Ser	Tyr	Ser 280	Phe	Ile	Arg	Lys	His 285	-	1105
40		Arg	Tyr	Ser 290	Lys	Lys	Thr	Ala	Сув 295	Val	Leu	Pro	Ala	Pro 300	Ala	GGA Gly	1153
45	Pro	Ser	Gln 305	Gly	Lys	His	Leu	Ala 310	Val	Pro	Glu	Asn	Pro 315	Ala	Ser	GTC Val	1201
73	Arg	Ser 320	Gln	Leu	Ser	Pro	Ser 325	Ser	Lys	Val	Ile	Pro 330	Gly	Val	Pro	Ile	
50	Сув 335	Phe	Glu	Val	Lys	Pro 340	Glu	Glu	Ser	Ser	Авр 345	Ala	His	Glu	Met	AGA Arg 350	1297
55											Arg					TTC Phe	1345

PCT/US97/05781 WO 97/37998

	TAC AGA CTG ACC ATA CTG ATA CTC GTG TTC GCC GTT AGC TGG ATG CCA	1393
	Tyr Arg Leu Thr Ile Leu Ile Leu Val Phe Ala Val Ser Trp Met Pro 370 375 380	
5	CTC CAC GTC TTC CAC GTG GTG ACT GAC TTC AAT GAT AAC TTG ATT TCC Leu His Val Phe His Val Val Thr Asp Phe Asn Asp Asn Leu Ile Ser	1441
	385 390 395 ANT AGG CAT TTC AAG CTG GTA TAC TGC ATC TGT CAC TTG TTA GGC ATG	1489
10	Asn Arg His Phe Lys Leu Val Tyr Cys Ile Cys His Leu Leu Gly Met 400 405 410	
	ATG TCC TGT TGT CTA AAT CCG ATC CTA TAT GGT TTC CTT AAT AAT GGT Met Ser Cys Cys Leu Asn Pro Ile Leu Tyr Gly Phe Leu Asn Asn Gly 415 420 425 430	1537
15	ATC AAA GCA GAC TTG AGA GCC CTT ATC CAC TGC CTA CAC ATG TCA TGA Ile Lys Ala Asp Leu Arg Ala Leu Ile His Cys Leu His Met Ser * 435 440 445	1585
20	TTCTCTCTGT GCACCAAAGA GAGAAGAAAC GTGGTAATTG ACACATAATT TATACAGAAG	1645
	TATTCTGGAT CTGAATGCCA GTTCGTAATC TACGTAAGAT CATCTTCATG TTATAATATG	1705
25	GTTAATTCAA TCAGTTGTGC AGAGTCAATG TCCATCTAAT ACAATTTCAT GTGTTGAAGT	1765
	AGTTTACATT ATTITCCATT TTATGTCATT GGTAATAAGT TGAGTGATAC TCTGTGGTTT	1825
	AGTGTAAAAG ATATAGCTAT CCAAATTGTT ACGTTGTACA AAAAATGTAT GAAGTGACAA	1985
30	GTTGTCCCAA AGAGCATTTA ACTACAGATT TAAGGAATTT CTATTATCTG GGTATCTTCA TTTCTATTTC ACAGGCTTCT TAACATTTTT TTGTAAAAGT ACAAAAATAT TCAAAAGTCA	2005
	GAACTCTATT ACAGATGTAT GCATAAAAGA TGATTATAAT TTTGTAGGAG AAAGATCTGC	2065
35	TCCTATTAGT GAAGATTGGT AAAATTGTCA GTTTAACCCG GCTGTCCTAC TACTAATATT	2125
	TAATTITTCA AATATGAAAA GGTTTCAGAT TTTGTTTAGA TITATATCAC ATTAAACACT	2185
40	GTCAAATAAA GGCTGTTTTT ATATGCATCG TTGATGTTCC AAAATGTGAA GTCTAAATGG	2245
	TGTCTGTATT TCCAATTATT AAATAACTTC TAAGATCATT TTTAAAAGTC TGTAGATGGT	2305
45	ATGGATAGCT AGTTGTTTGT TAATATAAAG TAAAAGTAGA TAGCTGATTT ATGTTGTACC	2365
	TATGTCGTAT GTATATTAGG TATCGTGTTG TCTCACTAAA GTGAAAGCAA ACGAAAAAAA	2425
	AAAAAA AAAAAAAAA AAAAAAAAA AAAAAAAAA AAAA	2481

(2) INFORMATION FOR SEQ ID NO:2:

50

55

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 445 amino acids

 - (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Glu Phe Lys Leu Glu Glu His Phe Asn Lys Thr Phe Val Thr Glu . Asn Asn Thr Ala Ala Ala Arg Asn Ala Ala Phe Pro Ala Trp Glu Asp 25 Tyr Arg Gly Ser Val Asp Asp Leu Gln Tyr Phe Leu Ile Gly Leu Tyr 10 Thr Phe Val Ser Leu Leu Gly Phe Met Gly Asn Leu Pro Ile Leu Met 15 Ala Val Met Lys Lys Arg Asn Gln Lys Thr Thr Val Asn Phe Leu Ile Gly Asn Leu Ala Phe Ser Asp Ile Leu Val Val Leu Phe Cys Ser Pro 20 Phe Thr Leu Thr Ser Val Leu Leu Asp Gln Trp Met Phe Gly Lys Ala 25 Met Cys His Ile Met Pro Phe Leu Gln Cys Val Ser Val Leu Val Ser Thr Leu Ile Leu Ile Ser Ile Ala Ile Val Arg Tyr His Met Ile Lys 130 135 30 His Pro Ile Ser Asn Asn Leu Thr Ala Asn His Gly Tyr Phe Leu Ile 150 Ala Thr Val Trp Thr Leu Gly Phe Ala Ile Cys Ser Pro Leu Pro Val 35 Phe His Ser Leu Val Glu Leu Lys Glu Thr Phe Gly Ser Ala Leu Leu 40 Ser Ser Lys Tyr Leu Cys Val Glu Ser Trp Pro Ser Asp Ser Tyr Arg Ile Ala Phe Thr Ile Ser Leu Leu Val Gln Tyr Ile Leu Pro Leu 45 Val Cys Leu Thr Val Ser His Thr Ser Val Cys Arg Ser Ile Ser Cys Gly Leu Ser His Lys Glu Asn Arg Leu Glu Glu Asn Glu Met Ile Asn 50 245 Leu Thr Leu Gln Pro Ser Lys Lys Ser Arg Asn Gln Ala Lys Thr Pro 265 55 Ser Thr Gln Lys Trp Ser Tyr Ser Phe Ile Arg Lys His Arg Arg Arg 280 275

	Tyr	Ser 290	Lys	Lys	Thr	Ala	Cys 295	Val	Leu	Pro	Ala	9ro 300	Ala	GIA	Pro	Ser
5	Gln 305	Gly	Lys	His	Leu	Ala 310	Val	Pro	Glu	Asn	Pro 315	Ala	Ser	Val	Arg	Ser 320
	Gln	Leu	Ser	Pro	Ser 325	Ser	ГЛа	Val	Ile	Pro 330	Gly	Val	Pro	Ile	Cys 335	Phe
10	Glu	Val	Lys	Pro	Glu	Glu	Ser	Ser	Asp 345	Ala	His	Glu	Met	Arg 350	Val	Lys
	Arg	Ser	Ile 355	Thr	Arg	Ile	Lys	360	Arg	Ser	Arg	Ser	Val 365	Phe	Tyr	Arg
15	Leu	Thr 370		Leu	Ile	Leu	Val 375		Ala	Val	Ser	Trp 380	Met	Pro	Leu	His
20	Val 385		His	Val	Val	Thr 390		Phe	. Asn	Asp	Asn 395	Leu	Ile	Ser	Asn	Arg 400
	His	Phe	Lys	Leu	Val 405		Сув	Ile	: Сув	His 410	Leu)	Leu	Gly	Met	415	Ser
25	Сув	Сув	Lev	420		Ile	Lev	туг	Gly 425	Phe	Lev	a Asn	Asr	430	/ Ile	e Lys
30	Ala	y yat	435		, A la	a Leu	ı Ile	His		3 Leu	ı His	Met	Se:	s * 5		
30	(2)	INE	FORM	ATIO	N FOI	R SE	Q ID	NO:	3:							
35	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2604 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear															
40						TYPE		AN								
		•				E: N										
45		(i	.x) F		NAME	E/KEY			1708							
50		()	L x) 1		MAM	E/KET				de						
55		1-	xi)	SEOU	ENCE	DES	CRIP	TION	: SE	Q ID	NO:	3:				

32

	GGGA	.ctgt	CA C	GTGT	TCCC	G AG	GTGC	TTCT	AAA	ACCC	TGG	CGGC	rccg	ga g	cccc	ICCII	120
	CCCA	CCAC	cg c	CTCC	aggt	c cr	GCTC	CTGC	CGC	CACC	GCT	TCCA	TCTG	GA G	CAGA	AGCGA	180
5	CCGC	GCTC	AG C	CACG	TACC	C CG	GAGT	CCAG	GCA	CCCG	CAG	CGGC	CGGG	GC A	TCCC	GAGCT	240
	GGCC	ATAC	AC C	GGGA	GACA	G CT	GTGC	CCTT	GGG	TTTG	CAA	GGTG	GCTT	GG A	AGTC	AACTG	300
10	CCAG	TAGG	SAA A	TAGC	CATO	C AC	ACAC	CTGA	GTT	CCAA	.GGG	GGAA	GAAA	ga g	ATTC	TTATC	360
	TGATTTTAGT ATG GAG TTT AAG CTT GAG GAG CAT TTT AAC AAG ACA TTT Met Glu Phe Lys Leu Glu Glu His Phe Asn Lys Thr Phe 1 5 10															409	
15			GAG Glu														457
20			GAC Asp														505
25			TAT Tyr														553
30			ATG Met														601
30			ATA Ile 80														649
35			CCT Pro														697
40		Lys	GCC Ala														745
45			TCA Ser								Ala					CAT	793
50			AAG Lys							Leu					Gly	TAC Tyr	841
			ATA Ile 160	Ala					Leu					САа		CCC Pro	889
55			Val					Val					Thr			TCA Ser	937

	GCA Ala 190	CTG Leu	CTG Leu	AGT Ser	Ser	AAA ' Lys ' 195	IAT (CTC '	TGT ·	Val	GAG Glu 200	TCA Ser	TGG Trp	CCC Pro	TCT Ser	GAT Asp 205		985
5	TCA Ser	TAC Tyr	AGA Arg	Ile	GCT Ala 210	TTC . Phe	ACA I	ATC Ile	TCT Ser	TTA Leu 215	TTG Leu	CTA Leu	GTG Val	CAG Gln	TAT Tyr 220	ATC Ile		1033
10	Leu	Pro	Leu	Val 225	Сув	TTA Leu	Thr	Val	Ser 230	His	Thr	Ser	Val	Cys 235	Arg	Ser		1081
15	lle	Ser	Сув 240	Gly	Leu		His	Lys 245	Glu	Asn	Arg	Leu	Glu 250	Glu	Asn	Glu		
	ATG Met	ATC Ile 255	AAC Asn	TTA Leu	ACC Thr	CTA Leu	CAG Gln 260	CCA Pro	TCC Ser	AAA Lys	Lys Lys	AGC Ser 265	AGG Arg	AAC Asn	Gln	GCA Ala	•	
20	AAA Lys 270	ACC Thr	CCC	AGC Ser	ACT Thr	CAA Gln 275	AAG Lys	TGG Trp	AGC Ser	TAC Tyr	TCA Ser 280	Phe	ATC Ile	AGA Arg	Lys	CAC His 285	3	1225
25	AGA Arg	AGG Arg	AGG Arg	TAC Tyr	AGC Ser 290	AAG Lys	AAG Lув	ACG Thr	GCC Ala	ТGТ Сув 295	Val	TTA Leu	CCC Pro	GCC Ala	Pro 300	Ala		1273
30	GGA Gly	CCT	Ser	CAG Gln 305	Gly	T Aa	CAC	CTA Leu	GCC Ala 310	Val	CCA Pro	GAA Glu	AAT Asn	Pro 315	Ala	C TCC	c r	1321
35	GTC Val	CGT	AGC Ser 320	Gln	CTG Leu	TCG Ser	CCA Pro	TCC Ser 325	Ser	AAG Lys	GTC Val	ATI Ile	CCA Pro	Gly	GT Va	c cci	A O	1369
40	ATC Ile	TG0	Phe	GAG	GTG Val	Lys	Pro	Glu	GAA	AGC Ser	Ser	A GAT c Asp 34!	p Ala	r CAT	r GA s Gl	G AT u Me	G t	1417
40	AGA Arg 350	Va:	L Lyi	G CGT	TCC Ser	: ATC : Ile :355	Thr	AGA	ATA J Ile	AAJ Lys	A AAG E Ly: 36	s Ar	A TC	r CG.	A AG g Se	T GT T Va 36	11	1465
45	TTC Phe	TA	C AG	A CTO	3 ACC 1 Thi 370	C ATA	CTG Leu	ATA Ile	CT(GT(1 Va) 37	l Ph	C GC e Al	C GT a Va	T AG l Se	C TO T T1	TO Me	r G et	1513
50	Pro	A CT	C CA u Hi	C GT(B Va:	l Pho	C CAC	GTC Val	GT(G AC 1 Th 39	r As	C TT P Ph	C AA le As	T GA n As	T AA p As 39	n L	rg A1 eu Il	rr le	1561
55	TC: Se:	C AA	T AG n Ar 40	g Hi	T TT	C AAG e Ly	G CT(G GT. u Va 40	l Ty	C TG T Cy	ra Il	C TO	ST CA 78 Hi 41	s Le	G T	TA GO	GC ly	1609

	ATG ATG TCC TGT TGT CTA AAT CCG ATC CTA TAT GGT TTC CTT AAT AAT Met Met Ser Cys Cys Leu Asn Pro Ile Leu Tyr Gly Phe Leu Asn Asn 415 420 425	1657
5	GGT ATC AAA GCA GAC TTG AGA GCC CTT ATC CAC TGC CTA CAC ATG TCA Gly Ile Lys Ala Asp Leu Arg Ala Leu Ile His Cys Leu His Met Ser 430 440 445	1705
10	TGA TTCTCTCTGT GCACCAAAGA GAGAAGAAAC GTGGTAATTG ACACATAATT	1758
	TATACAGAAG TATTCTGGAT CTGAATGCCA GTTCGTAATC TACGTAAGAT CATCTTCATG	1818
15	TTATAATATG GTTAATTCAA TCAGTTGTGC AGAGTCAATG TCCATCTAAT ACAATTTCAT	1878
	GTGTTGAAGT AGTTTACATT ATTTTCCATT TTATGTCATT GGTAATAAGT TGAGTGATAC	1938
20	TCTGTGGTTT AGTGTAAAAG ATATAGCTAT CCAAATTGTT ACGTTGTACA AAAAATGTAT	1998
	GAAGTGACAA GTTGTCCCAA AGAGCATTTA ACTACAGATT TAAGGAATTT CTATTATCTG	2058
	GGTATCTTCA TTTCTATTTC ACAGGCTTCT TAACATTTTT TTGTAAAAGT ACAAAAATAT	2118
25	TCAAAAGTCA GAACTCTATT ACAGATGTAT GCATAAAAGA TGATTATAAT TTTGTAGGAG	2178
	AAAGATCTGC TCCTATTAGT GAAGATTGGT AAAATTGTCA GTTTAACCCG GCTGTCCTAC	2238
30	TACTAATATT TAATTTTTCA AATATGAAAA GGTTTCAGAT TTTGTTTAGA TTTATATCAC	2298
	ATTAAACACT GTCAAATAAA GGCTGTTTTT ATATGCATCG TTGATGTTCC AAAATGTGAA	2358
	GTCTAAATGG TGTCTGTATT TCCAATTATT AAATAACTTC TAAGATCATT TTTAAAAGTC	2418
35	TGTAGATGGT ATGGATAGCT AGTTGTTTGT TAATATAAAG TAAAAGTAGA TAGCTGATTT	2478
	ATGTTGTACC TATGTCGTAT GTATATTAGG TATCGTGTTG TCTCACTAAA GTGAAAGCAA	2538
40	ACGAAAAAA AAAAAAAAA AAAAAAAAA AAAAAAAAA AAAA	2598
	AAAAA	2604

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

45

50

- (A) LENGTH: 445 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:
- 55 Met Glu Phe Lys Leu Glu Glu His Phe Asn Lys Thr Phe Val Thr Glu

 1 5 10 15

	Asn	Asn	Thr	Ala . 20	Ala	Ala	Arg	Asn	Ala . 25	Ala	Phe	Pro /	Ala	Trp 30	Glu	Asp
5	Tyr	Arg	Gly 35	Ser	Val	Asp	Asp	Leu 40	Gln	Tyr	Phe	Leu	Ile 45	Gly	Leu	Tyr
	Thr	Phe 50	Val	Ser	Leu	Leu	Gly 55	Phe	Met	Gly _.	Asn	Leu 60	Leu	Ile	Leu	Met
10	Ala 65	Val	Met	Lys	Lys	Arg 70	Asn	Gln	Lys	Thr	Thr 75	Val	Asn	Phe	Leu	Ile 80
	Gly	Asn	Leu	Ala	Phe 85	Ser	qaA	Ile	Leu	Val 90	Val	Leu	Phe	Суз	Ser 95	Pro
15	Phe	Thr	Leu	Thr 100	Ser	Val	Leu	Leu	Asp 105	Gln	Trp	Met	Phe	Gly 110	Lys	Ala
20	Met	Сув	His 115	Ile	Met	Pro	Phe	Leu 120	Gln	Cys	Val	Ser	Val 125	Leu	Val	Ser
	Thr	Leu 130		Leu	Ile	Ser	Ile 135	Ala	Ile	Val	Arg	Tyr 140	His	Met	Ile	Lys
25	His 145	Pro	Ile	Ser	Asn	Asn 150	Leu	Thr	Ala	Asn	His 155	Gly	Tyr	Phe	Leu	Ile 160
70	Ala	Thr	Val	Trp	Thr 165		Gly	Phe	Ala	Ile 170		Ser	Pro	Leu	Pro 175	Val
30	Phe	His	Ser	Leu 180	Val	Glu	Leu	Lys	Glu 185		Phe	Gly	Ser	Ala 190	Leu	Leu
35·	Ser	Ser	Lys 195		Leu	Cys	Val	Glu 200		Trp	Pro	Ser	Asp 205	Ser	туг	Arg
40	Ile	210		. Thr	Ile	. Ser	Leu 215		ı Lev	val	. Gln	Tyr 220	: Ile	e Leu	ı Pro) Leu
40	Va] 225		. Lev	ı Thr	Val	230		3 Thi	r Ser	· Val	235		g Ser	r Ile	a Se	r Cys 240
45	Gly	y Lei	ı Se	r His	Lys 245		IEA L	n Arg	g Lev	250		ı Ası	n Gli	u Mei	25	e Asn 5
	Lei	u Th	r Le	u Glr 260		o Se	r Ly:	s Ly	8 Se:		g Ası	n Gl	n Al	a Ly 27	s Th O	r Pro
50	Se	r Th	r Gl 27		Tr	p Se	т ту	r Se 28		e Il	e Ar	g Ly	s Hi 28	s Ar 5	g Ar	g Arg
5.5	Тy	r Se 29		s Ly	s Th	r Al	а Су 29		l Le	u Pr	o Al	a Pr 30	0 Al	a Gl	y Pr	o Ser
55	G1 30		у Lу	e Hi	s Le	u Al 31		l Pr	o Gl	u As	n Pr 31	o Al	a Se	r Va	il Ai	rg Sei 320

	Gln	Leu	Ser	Pro	Ser 325	Ser	Lys	Val	Ile	Pro 330	Gly	Val	Pro	Ile	335	Phe	
5	Glu	Val	Lys	Pro 340	Glu	Glu	Ser	Ser	Asp 345	Ala	His	Glu	Met	Arg 350	Val	Lys	
	Arg	Ser	Ile 355	Thr	Arg	Ile	Lys	L ув 360	Arg	Ser	Arg	Ser	Val 365	Phe	Tyr	Arg	
10	Leu	Thr 370	Ile	Leu	Ile	Leu	Val 375	Phe	Ala	Val	Ser	Trp 380	Met	Pro	Leu	His	
15	Val 385	Phe	His	Val	Val	Thr 390	Ąsp	Phe	Asn	Asp	Asn 395	Leu	Ile	Ser	Asn	Arg 400	
	His	Phe	Lys	Leu	Val 405	Tyr	Cys	Ile	Суз	His 410	Leu	Leu	Gly	Met	Met 415	Ser	
20				420	Pro				425					430	Ile	Lys	
	Ala	Asp	Leu 435	Arg	Ala	Leu	Ile	His 440	Cys	Leu	His	Met	Ser 445	•	-		
25	(2)				FOR											-	
30		(1	(. (:	A) L B) T C) S	ENGT YPE: TRANI	H: 1 nuc DEDN	338 leic ESS:	base aci sin	pai: d	rs							
		(ii) MO:	LECU	LE T	YPE:	gen	omic	DNA								
35		(iii) HY	РОТН	ETIC	AL:	NO										
40) FE	ATUR	ense B: Ame/:			-									
		1 4	(B) L	OCAT												
45		(1%		A) N	e: Ame/ Ocat			_									
		(xi) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	0:5:						
50		Авр				Asp					Lys					GAG Glu	48
55					Ala					Asp			-		ABE	GAC	96

	TAT Tyr	AAA Lys	AGC Ser 35	AGT Ser	GTA Val	GAT Asp	GAC Asp	TTA Leu 40	CAG Gln	TAT Tyr	TTT Phe	CTG Leu	ATT Ile 45	GGG Gly	CTC Leu	TA Ty	T	14	44
5	ACA Thr	TTT Phe 50	GTA Val	AGT Ser	CTT Leu	CTT Leu	GGC Gly 55	TTT Phe	ATG Met	GGG Gly	Asn	CTA Leu 60	CTT Leu	ATT Ile	TTA Leu	AT Me	G :t	1	92
10	GCT Ala 65	CTC Leu	ATG Met	AAA Lys	AAG Lys	CGT Arg 70	TAA naA	CAG Gln	AAG Lys	ACT Thr	ACG Thr 75	GTA Val	AAC Asn	TTC Phe	CTC	11	ra Le Bo	2	40
15	GGC Gly	TAA Asn	CTG Leu	GCC Ala	TTT Phe 85	TCT Ser	GAT Asp	ATC Ile	TTG Leu	GTT Val 90	GTG Val	CTG Leu	TTT	TGC Cys	TCA Ser 95	P	et ro	2	88
30:	TTC Phe	ACA Thr	CTG Leu	ACG Thr 100	TCT Ser	GTC Val	TTG Leu	CTG Leu	GAT Asp 105	CAG Gln	TGG Trp	ATG Met	TTT Phe	GGC Gly 110	Lys	A G	TC al	3	336
20	ATG Met	TGC Cys	CAT His 115	ATT Ile	ATG Met	CCT Pro	TTT Phe	CTT Leu 120	CAA Gln	TGT Cys	GTG Val	TCA Ser	GTT Val 125	Leu	GT1	r T L S	CA er	:	384
25	ACT Thr	TTA Leu 130	Ile	TTA Leu	ATA Ile	TCA Ser	ATT Ile 135	GCC Ala	ATT	GTC Val	AGG Arg	TAT Tyr 140	His	Met	AT	A A e L	AA ys		432
30	CAT His 145	Pro	ATA	TCT Ser	TAA '	AAT Asn 150	Leu	ACA Thr	GCA Ala	AAC nea	CAT His	Gly	TAC Tyl	Phe	CT Le	u I	TA le 160		480
35	GCT Ala	ACI Thi	GTC Val	TGG	ACA Thr	Leu	GGT	TTT Phe	GCC Ala	116 170	Cys	r TCT	r CCC	CT.	r cc u Pr 17	0 1	STG Val		528
40	TTT	CAC Hi	E AG	r CTT	ı Val	G GAA	CTI Let	CAA Glr	GAF Glu 185	Thi	TT:	r GG e Gl	T TC. y Se	A GC r Al 19	a Le	G (CTG Leu		5 76
40	AG(Se)	Z AG	C AGG	G TAT g Ty: 5	r TT	A TGT	r GTT 3 Val	GAC Glu 200	ı se	A TGG	CC.	A TC o Se	T GA r As 20	p Se	A TA	AC yr	AGA Arg		624
45	AT.	T GC e Al 21	a Ph	T AC	T AT	C TC	TT: r Let 21:	ı Lev	G CT	A GT u Va	r CA 1 Gl	G TA n Ty 22	r Il	T CI	G C u P:	cc	TTA Leu		672
50	GT Va 22	1 су	T CT	T AC	T GT r Va	A AG 1 Se 23	r Hi	T AC	A AG r Se	T GT r Va	C TG 1 Cy 23	rs Ar	GA AC	T AT	ra a Le s	GC er	TGT Cys 240		720
55	GG G1	A TI	G TO	C AA	C AA n Ly 24	s Gl	A AA u As	C AG n Ar	A CT	T GA :u Gl 25	.u G]	AA AI Lu Ai	AT GI sn Gi	AG A' Lu M	et I	TC le	AAC Asn		768

			•															
	TTA	ACT	CTT	CAT	CCA	TCC	AAA	AAG	AGT	GGG	CCT	CAG	GTG	AAA	CTC	TCT		816
	Leu	Thr	Leu	His	Pro	Ser	Lys	Lys	Ser	Gly	Pro	Gln	Val	Lys	Leu	Ser		
				260			-		265					270				
5	GGC	AGC	CAT	AAA	TGG	AGT	TAT	TCA	TTC	ATC	AAA	AAA	CAC	AGA	AGA	AGA		864
_	Glv	Ser	His	Lvs	Trp	Ser	Tyr	Ser	Phe	Ile	Lys	Lys	His	Arg	Arg	Arg		
	,		275	•	•		•	280					285					
	TAT	AGC	AAG	AAG	ACA	GCA	TGT	GTG	TTA	CCT	GCT	CCA	GAA	AGA	CCT	TCT		912
10	Tvr	Ser	Lvs	Lvs	Thr	Ala	Cvs	Val	Leu	Pro	Ala	Pro	Glu	Arg	Pro	Ser		
-	- 4 -	290		-4 -			295					300						
	CAA	GAG	AAC	CAC	TCC	AGA	ATA	CIT	CCA	GAA	AAC	TTT	GGC	TCT	GTA	AGA		960
	Gln	Glu	Agn	His	Ser	Arg	Tle	Leu	Pro	Glu	Asn	Phe	Gly	Ser	Val	Arq		
15	305	314	70	*****	001	310					315		1			320		
	505					710												
	ACT	CAG	CTC	тст	TCA	TCC	ACT	DAG	TTC	ATA	CCA	GGG	GTC	CCC	ACT	TGC		1008
						Ser												
	361	GIII	Deu	Jer	325	J C.	562	2,5		330		02,			335	-,		
20	444	CNG	מדה	מממ'		GAA	CAA	ТАА	TCA		GTT	ТДЭ	GAA	TTG		ATD.		1056
20						Glu												1030
	PILE	GIU	116	340	PIU	GIU	GIU	ASII	345	vob	V 41	*****	014	350		***		
				340					343					330				
	מממ	CCT	тст	CTT	חכח	D.C.D.	מדה	222	AAG	AGA	TCT	CGA	AGT	GTT	רדדר	TAC		1104
25																Tyr	•	
23	Dyb	AL 9	355		1111	Arg	116	360	Lyb	719	Jer	71.9	365			- , -		
			333					300					303					
	»C»	CTG	» CC	ስ T ከ	CTC:	2.72	עידשר	CTA	TTT	CCT	تحت	AGT	TCC	ATG	CCA	CTA		1152
																Leu		
30	AL 9	370	1111	116	Leu	116	375	VAI	FIIC	, L.	vai	380	1-1			200		
50		3,0					3,7					300						
	CAC	СТТ	كملمك	САТ	GTG	CTA	۸ ۲۰۰۰	CAT	للململة	ТАА	GAC	דמג	СТТ	יידע	TCA	AAT		1200
																Asn		
	385		•		• • • •	390					395					400		
35	555					3,0												
	AGG	САТ	77	DAG	TTC	GTG	TAT	TGC	ATT	тст	САТ	ттс	TTG	GGC	ATG	ATG		1248
																Met		
	9			Lys	405		- 7 -	Cys		410		200		,	415			
					403													
40	TCC	TGT	TGT	СТТ	AAT	CCA	ATT	СТА	TAT	GGG	T-T-T	СТТ	AAT	` AAT	GGG	ATT		1296
																Ile		
		٠,٥	-,5						425			200	, ,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	430				
				-20														
45	AAA	GCT	GAT	מידן	GTG	TCC	CTT	ATA	CAC	TGT	СТТ	CAT	ATO	TAR				1338
						Ser									-			
	-,-		435					440		-,-			445				•	
														-				

- 50 (2) INFORMATION FOR SEQ ID NO:6:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 445 amino acids
 - (B) TYPE: amino acid
- 55 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein

(xi)	SEQUENCE	DESCRIPTION:	SEQ	ID	NO:6:
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	Met 1	Asp	Leu	Glu	Leu 5	Asp	Glu	Tyr	Tyr	Asn 10	Lys	Thr	Leu	Ala	Thr 15	Glu
5	Asn	Asn	Thr	Ala 20	Ala	Thr	Arg	Asn	Ser 25	Asp	Phe	Pro	Val	Trp 30	Asp	qaA
10	туг	Lys	Ser 35	Ser	Val	Asp	Asp	Leu 40	Gln	Tyr	Phe	Leu	Ile 45	Gly	Leu	Tyr
	Thr	Phe 50	Val	Ser	Leu	Leu	Gly 55	Phe	Met	Gly	naA	Leu 60	Leu	Ile	Leu	Met
15	Ala 65	Leu	Met	Lув	Lys	Arg 70	Asn	Gln	Lys	Thr	Thr 75	Val	Asn	Phe	Leu	Ile 80
20	Gly	Asn	Leu	Ala	Phe 85	Ser	Asp	Ile	Leu	Val 90	Val	Leu	Phe	Сув	Ser 95	Pro
20	Phe	Thr	Leu	Thr 100	Ser	Val	Leu	Leu	Asp 105	Gln	Trp	Met	Phe	Gly 110	Lys	Val
25	Met	Сув	His 115	Ile	Met	Pro	Phe	Leu 120	Gln	Сув	Val	Ser	Val 125	Leu	Val	Ser
	Thr	Leu 130	Ile	Leu	Ile	Ser	Ile 135	Ala	Ile	Val	Arg	Tyr 140	His	Met	Ile	Lys
30	His 145		Ile	Ser	Asn	Aśn 150		Thr	Ala	лал	His 155		Tyr	Phe	Leu	Ile 160
35	Ala	Thr	Val	Trp	Thr 165		Gly	Phe	Ala	Ile 170		Ser	Pro	Leu	Pro 175	Val
33	Phe	His	Ser	Leu 180		. Glu	Leu	Gln	Glu 185		Phe	Gly	Ser	Ala 190	Leu	Leu
40	Ser	Ser	195		Leu	Cys	Val	. Glu 200		Trp	Pro	Ser	205		Туг	Arg
	Ile	210		Thr	Ile	e Ser	215		Lev	ı Val	l Glr	220		e Lei	Pro	Leu
45	Va] 225		Let	ı Thr	· Val	230		Thi	Ser	· Val	235		g Sei	r Ile	e Se	т Сув 240
50	Gl	y Lev	ı Se:	r Ası	1 Ly:		ı Ası	n Arg	g Lev	25°		ı Ası	n Gli	u Mei	25	e Asn 5
30	Let	u Thi	r Le	u His 260		o Se	r Ly	s Ly	s Se:		y Pr	o Gl	n Va	1 Ly 27	в Le O	u Ser
55	G1	y Se:	r Hi 27		s Tr	p Se	r Ty	r Se 28		e Il	e Ly	s Ly	8 Hi 28	s Ar 5	g Ar	g Arg

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

	Met 1	Asp	Leu	Glu	Leu 5	Asp	Glu	Tyr "	Tyr .	Asn 10	Lys	Thr	Leu	Ala	Thr 15	Glu
5	Asn	naA	Thr	Ala 20	Ala	Thr	Arg	Asn	Ser 25	Asp	Phe	Pro	Val	Trp 30	Asp	Asp .
10	Туг	Lys	Ser 35	Ser	Val	Asp	Asp	Leu 40	Gln	Tyr	Phe	Leu	Ile 45	Gly	Leu	Tyr
	Thr	Phe 50	Val	Ser	Leu	Leu	Gly 55	Phe	Met	Gly	naA	Leu 60	Leu	Ile	Leu	Met
15	65			Lув		70					75					80
20				Ala	85					90					95	
				Thr 100					105					110		
25			115					120					125			Ser
		130					135					140				Lys
30	145				-	150	•				155					11e
35					165					170)				175	
				180					185					190)	. Leu
40			195	5				200	•				20	5		r Arg
		210)				215	5				220	0			. Leu
45	225	5				230)				23	5				r Cys 240
50					24	5				25	0				25	
-				260	0				26	5				27	0	u Sei
55	GJ.	y Se	r Hi 27		s Tr	p Se	т Ту	r Se: 28		e Il	e Ly	s Ly	's Hi 28	s Ar	g Ar	g Ar

	Tyr	Ser 290	Lys	Lys	Thr	Ala	Cys 295	Val	Leu	Pro	Ala	Pro 300	Glu	Arg	Pro	Ser		
5	Gln 305	Glu	Asn	His	Ser	Arg 310	Ile	Leu	Pro	Glu	Asn 315	Phe	Gly	Ser	Val	Arg 320		
-	Ser	Gln	Leu	Ser	Ser 325	Ser	Ser	Lys	Phe	Ile 330	Pro	Gly	Val	Pro	Thr 335	Cys		
10	Phe	Glu	Ile	Lys 340	Pro	Glu	Glu	Asn	Ser 345	Asp	Val	His	Glu	Leu 350	Arg	Val		
16	Lys	Arg	Ser 355	Val	Thr	Arg	Ile	Lys 360	Lys	Arg	Ser	Arg	Ser 365	Val	Phe	Tyr		
15	Arg	Leu 370	Thr	Ile	Leu	Ile	Leu 375	Val	Phe	Ala	Val	Ser 380	Trp	Met	Pro	Leu		
20	His 385	Leu	Phe	His	Val	Val 390	Thr	Asp	Phe	Asn	Asp 395	Asn	Leu	Ile	Ser	Asn 400		
	Arg	His	Phe	Lys	Leu 405	Val	Tyr	Cys	Ile	Cys 410	His	Leu	Leu	Gly	Met			
2 5	Ser	Cys	Сув	Leu 420		Pro	Ile	Leu	Tyr 425	Gly	Phe	Leu	Asn	Asn 430				
	Lys	Ala	Авр 435	Leu	Val	Ser	Leu	Ile 440	His	Cys	Leu	His	Met 445					
30	(2)	INF	ORMA	TION	FOR	SEQ	ID :	NO : 7	:									
35		(i	() ()	A) Li B) T C) S	ENGT: YPE : TRAN	H: 2 nuc	0 ba leic ESS:	ISTI se p aci sin ear	airs d							-		
		(ii) MO	LECU	LE T	YPE:	oth	er n	ucle	ic a	cid				•			
40																		
		(xi) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	0:7:							
45	TAG	GGAA	CCT	GGCC	TCCT	CC										٠	2	C
	(2)	INF	ORMA	TION	FOR	SEQ	ID	NO : 8	:									
50		(i	(. (.	A) L B) T C) S	ENGT YPE : TRAN	H: 2 nuc	0 ba leic ESS:	ISTI se p aci sin ear	airs d									
55		(ii) MO	LECU	LE T	YPE:	oth	er n	ucle	ic a	cid							

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:8:

TCAGAGGGCC ATGACTCAAC

20

We claim:

1. An isolated nucleic acid encoding a neuropeptide Y receptor comprising a molecule having a nucleotide sequence substantially the same as SEQ ID NO 1, SEQ ID NO 3, or SEQ ID NO 5.

- 2. A homogenous composition of the neuropeptide Y receptor comprising a molecule having an amino acid sequence substantially the same as SEQ ID NO 2, SEQ ID NO 4, or SEQ ID NO 6.
- 3. A vector comprising the nucleic acid according to claim 1.
- 4. A vector according to claim 3 adapted for expression in a cell further comprising regulatory elements necessary for expression of the nucleic acid operatively linked to the nucleic acid to enable expression of the nucleic acid.
- 5. A vector according to claim 4 wherein the cell is a mammalian cell.
- 6. A vector according to claim 5 wherein the cell is a human 293 cell.
- 7. A vector according to claim 4 that is a plasmid.
- 8. A vector according to claim 7 wherein the plasmid is the pBluescript plasmid.
- 9. A vector according to claim 7 wherein the plasmid is the pcDNA3 plasmid.
- 10. A vector according to claim 3 which is self-replicating.
- 11. A cell transformed with the nucleic acid according to claim 1 that expresses the nucleic acid.
- 12. A cell according to claim 11 that is a bacterium cell, an insect cell, or a yeast cell.
- 13. A cell according to claim 11 that is a mammalian cell.
- 14. A cell according to claim 13 that is a human 293 cell.

15. A nucleic acid probe comprising a nucleic acid complementary to the nucleic acid according to claim 1.

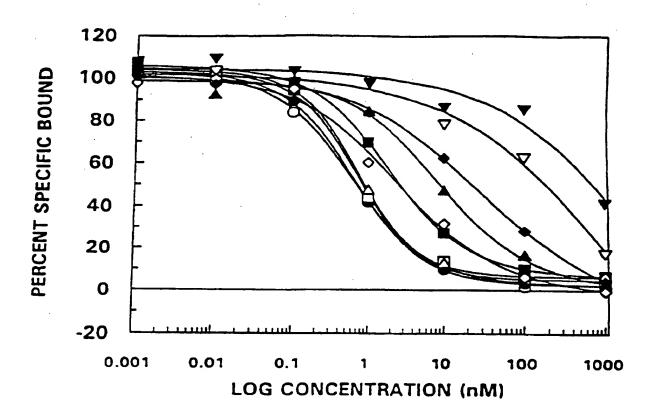
- 16. An antisense oligonucleotide having a sequence complementary to the nucleic acid according to claim 1 and that inhibits expression of the nucleic acid.
- 17. A membrane or membrane preparation comprising a membrane or portion thereof of a cell expressing a nucleic acid having a nucleotide sequence substantially the same as SEQ ID NO 1, SEQ ID NO 3, or SEQ ID NO 5.
- 18. A membrane or membrane preparation according to claim 17 wherein the cell is a mammalian cell.
- 19. An antibody or fragment thereof that is immunologically reactive to a mammalian Y5 receptor.
- 20. An antibody or fragment according to claim 19 thereof wherein the mammalian Y5 receptor has an amino acid sequence substantially the same as one chosen from the group consisting of SEQ ID NO 2, SEQ ID NO 4, and SEQ ID NO 6.
- 21. An antibody or fragment thereof according to claim 19 that is a monoclonal antibody.
- 22. An antibody or fragment thereof according to claim 20 that is a monoclonal antibody.
- 23. A cell line producing an antibody according to claim 19.
- 24. A cell line producing an antibody according to claim 20.
- 25. A cell line producing an antibody according to claim 21.
- 26. A cell line producing an antibody according to claim 22.
- 27. An epitope of a mammalian Y5 receptor wherein the epitope is immunologically reactive to the antibody or fragment thereof according to claim 19.
- 28. An epitope of a mammalian Y5 receptor wherein the epitope is immunologically reactive to the antibody or fragment thereof according to claim 20.

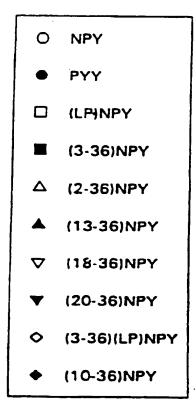
29. An epitope of a mammalian Y5 receptor wherein the epitope is immunologically reactive to the antibody or fragment thereof according to claim 21.

- 30. An epitope of a mammalian Y5 receptor wherein the epitope is immunologically reactive to the antibody or fragment thereof according to claim 22.
- 31. A method of producing a neuropeptide Y receptor comprising culturing a cell according to claim 11 and recovering the receptor expressed by the cell.
- 32. A method of producing a neuropeptide Y receptor comprising culturing a cell according to claim 12 and recovering the receptor expressed by the cell.
- 33. A method of producing a neuropeptide Y receptor comprising culturing a cell according to claim 13 and recovering the receptor expressed by the cell.
- 34. A method of producing a neuropeptide Y receptor comprising culturing a cell according to claim 14 and recovering the receptor expressed by the cell.
- 35. A method of identifying a neuropeptide Y agonist or antagonist comprising contacting a potential agonist or antagonist molecule with a membrane or membrane preparation according to claim 17.
- 36. A method of identifying a neuropeptide Y agonist or antagonist comprising contacting a potential agonist or antagonist molecule with a membrane or membrane preparation according to claim 18.
- 37. A neuropeptide Y antagonist comprising a compound identified according to claim 36.
- 38. A method of suppressing the appetite of a mammal comprising administering to the mammal an appetite suppressing amount of a neuropeptide Y antagonist according to claim 37.
- 39. A method of suppressing the appetite of a mammal according to claim 38 wherein the amount of antagonist is from about 0.01 to about 100 mg/kg.
- 40. A pharmaceutical composition comprising an effective appetite suppressing amount of an antagonist according to claim 37 together with a pharmaceutically acceptable carrier.

41. A neuropeptide Y agonist comprising a compound identified according to claim 36.

- 42. A method of stimulating the appetite of a mammal comprising administering to the mammal an appetite stimulating amount of a neuropeptide Y agonist according to claim 41.
- 43. A method of stimulating the appetite of a mammal according to claim 42 wherein the amount of agonist is from about 0.01 to about 100 mg/kg.
- 44. A pharmaceutical composition comprising an effective appetite stimulating amount of an agonist according to claim 41 together with a pharmaceutically acceptable carrier.
- 45. A non-human transgenic mammal that expresses the nucleic acid having a sequence substantially the same one chosen from the group consisting of SEQ ID NO 1, SEQ ID NO 3, and SEQ ID NO 5.





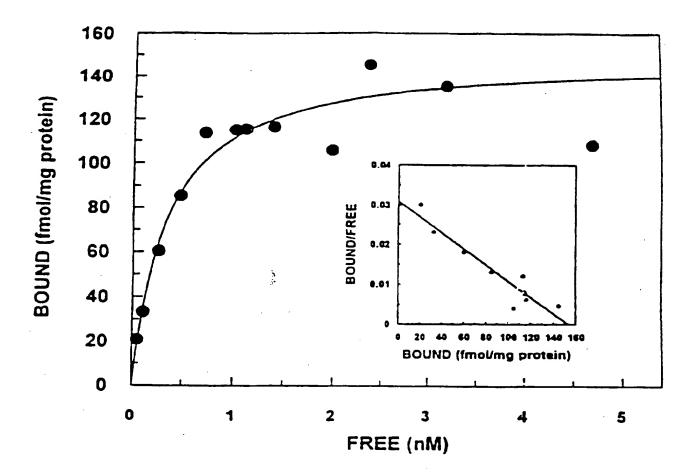


Fig. 2